

# Holographic Characterization of Protein Aggregates

Size, morphology and differentiation  
... one particle at a time (and fast)

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*New York University*

D. B. Ruffner & L.A. Philips  
*Spheryx, Inc*

# Holographic Characterization

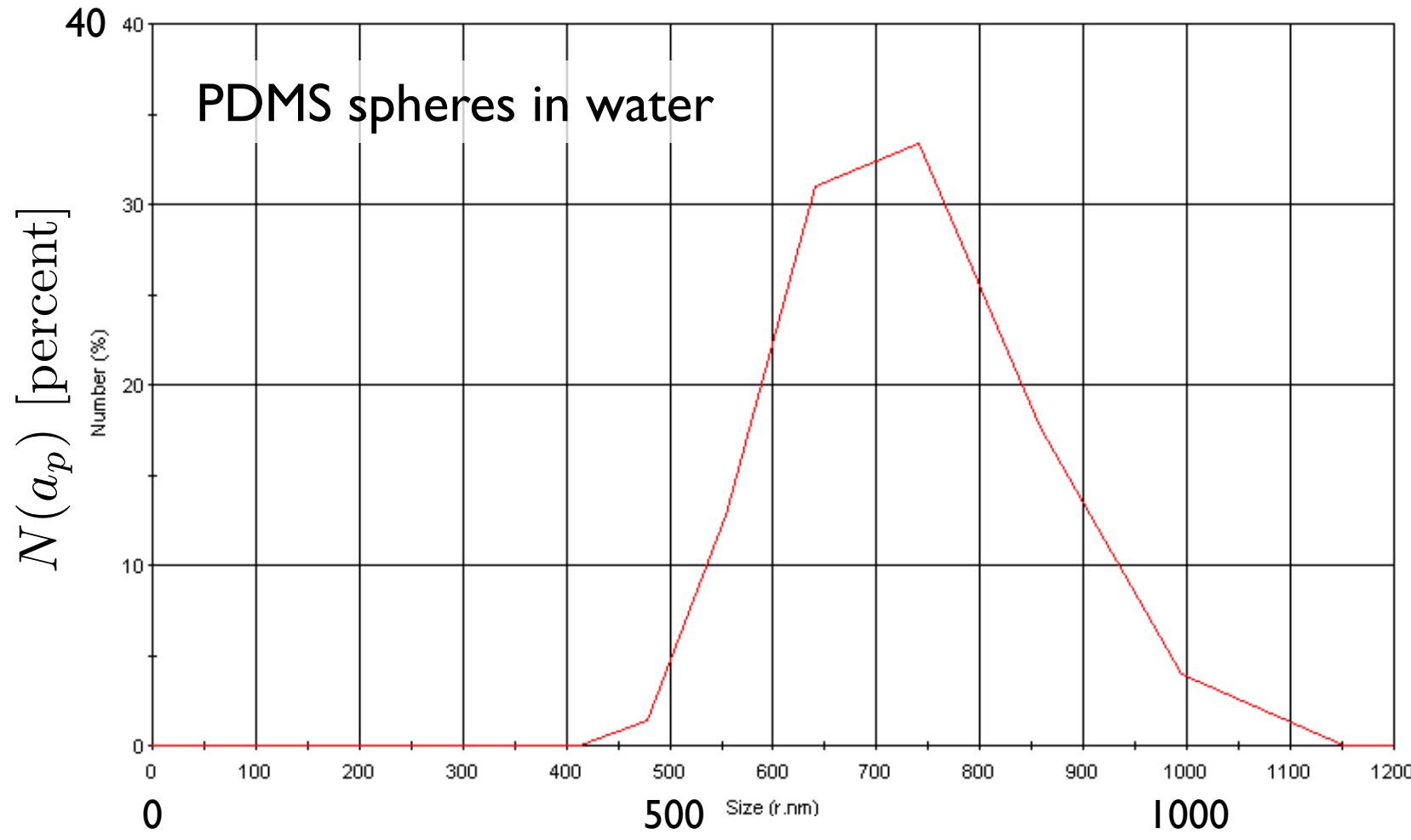
Particle-resolved measurements of

- **Radius:** subvisible range  
200 nm to 10  $\mu$ m
- **Refractive index:** proxy for composition  
Useful for differentiation
- **Morphology**  
Useful for differentiation  
Useful for analyzing aggregation mechanism
- **Concentration**  
Divide observed counts by measured sample volume
- **Real-time processes**  
30 measurements/second  
Complete analysis in 10 minutes

# So, what's the problem?



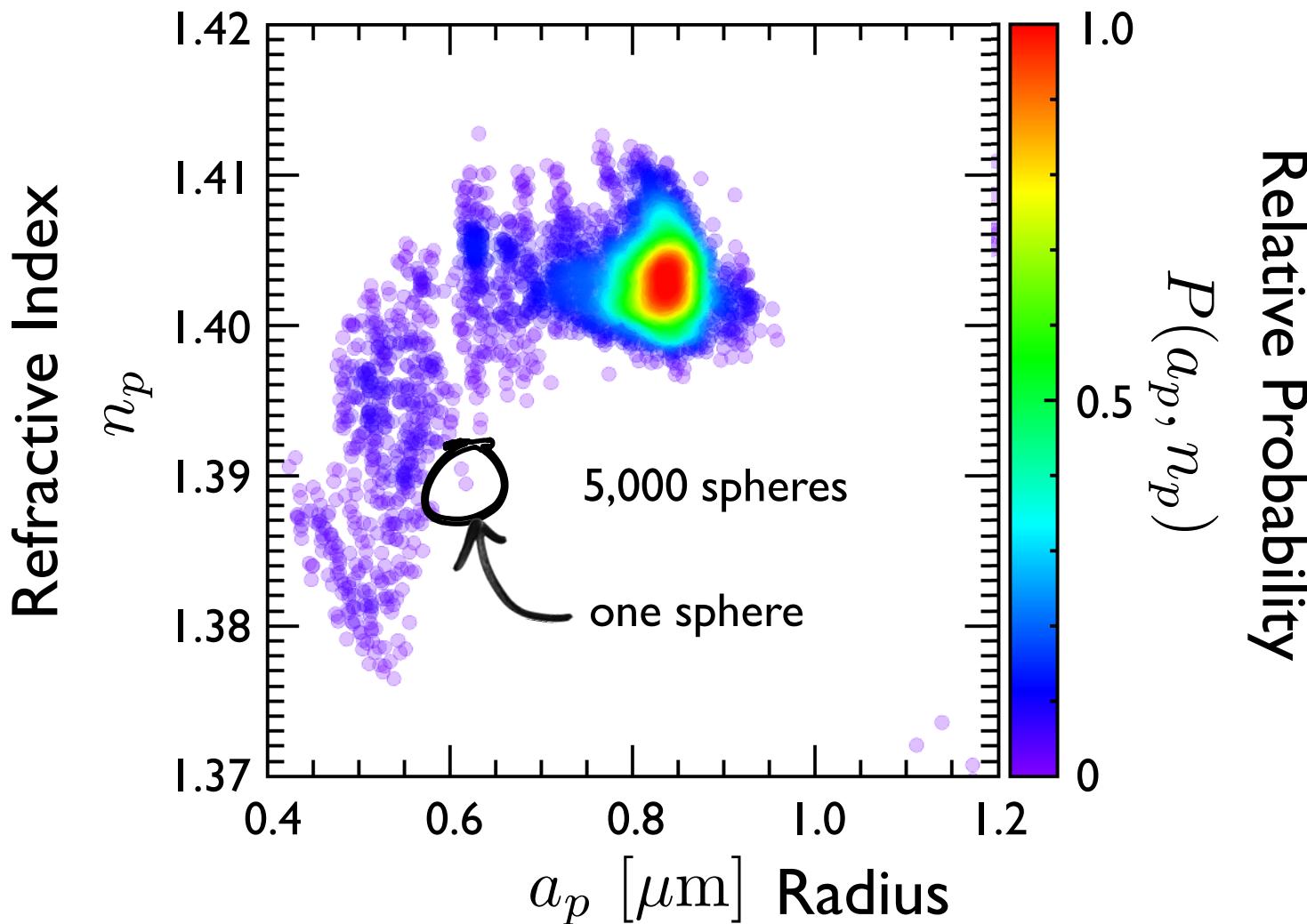
# Conventional Characterization (DLS)



Malvern Zetasizer Nano ZS

$a_p$  [nm]

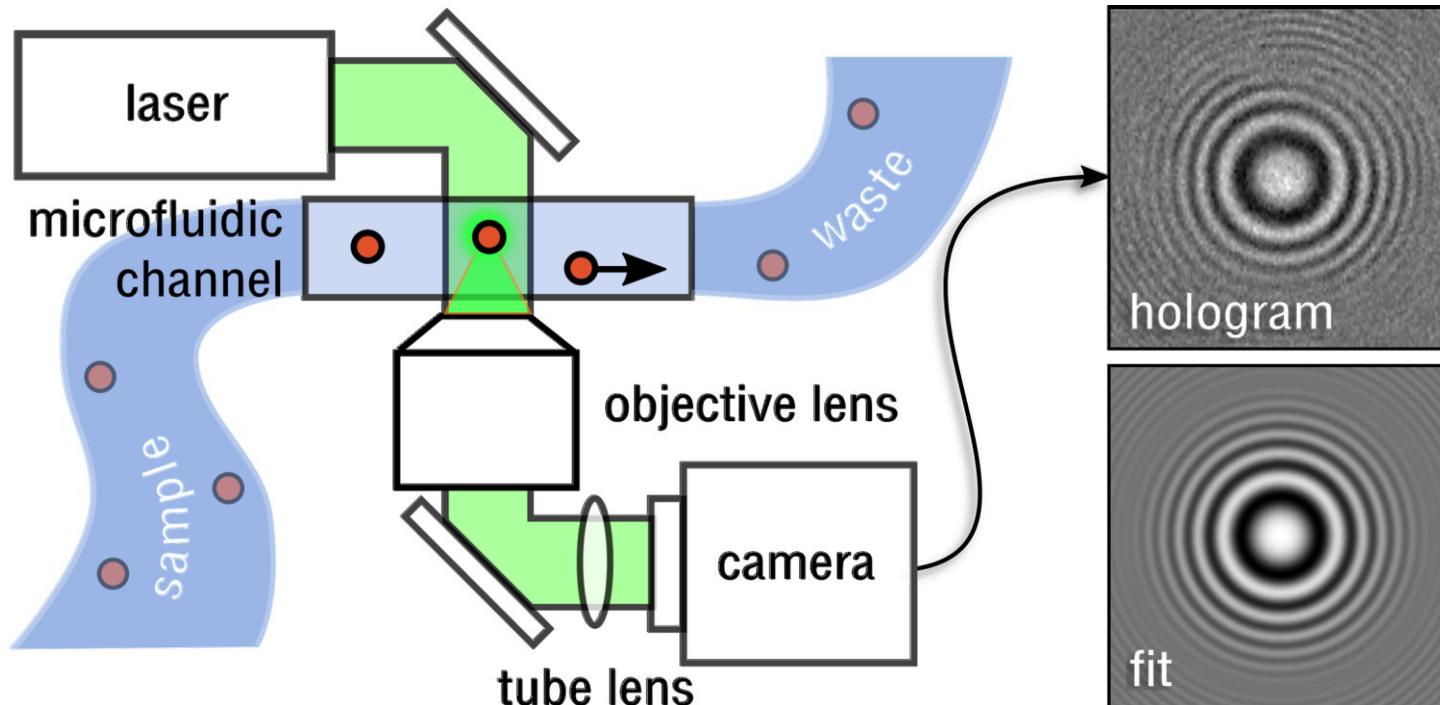
# Holographic Characterization



Wang, et al., Soft Matter 11, 1062 (2015)

# Holographic Tracking & Characterization

## Lorenz-Mie Microscopy

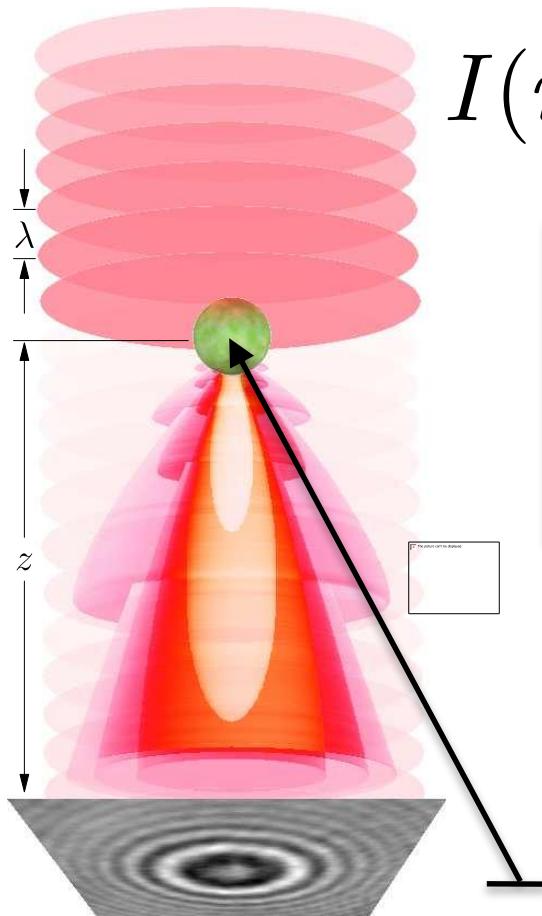


Parameters		
Tracking	3D Position	$r_p(t)$
Characterization	Radius	$a_p$
	Refractive Index	$n_p$

Lee et al., Optics Express 15, 18275 (2007)

# Holographic Tracking & Characterization

## How Lorenz-Mie Microscopy works



$$I(\mathbf{r}) = |E_0 + E_s(k(\mathbf{r} - \mathbf{r}_p))|^2$$

Incident plane wave:

$$k = \frac{2\pi n_m}{\lambda}$$

Scattered field:

$\mathbf{r}_p$  : position

$a_p$  : radius

$n_p$  : refractive index

Focal plane

# Holographic Tracking & Characterization

## How Lorenz-Mie Microscopy works

$$E_s(kr) = E_0 \sum_{n=1}^{\infty} i^n \frac{2n+1}{n(n+1)} \left( i a_n \mathbf{N}_{e1n}^{(3)}(kr) - b_n \mathbf{M}_{o1n}^{(3)}(kr) \right)$$
$$\mathbf{M}_{o1n}^{(3)}(kr) = \frac{\cos \phi}{\sin \theta} P_n^1(\cos \theta) j_n(kr) \hat{\theta}$$
$$- \sin \phi \frac{dP_n^1(\cos \theta)}{d\theta} j_n(kr) \hat{\phi}$$
$$\mathbf{N}_{e1n}^{(3)}(kr) = n(n+1) \cos \phi P_n^1(\cos \theta) \frac{j_n(kr)}{kr} \hat{r}$$
$$+ \cos \phi \frac{dP_n^1(\cos \theta)}{d\theta} \frac{1}{kr} \frac{d}{dr} [r j_n(kr)] \hat{\theta}$$
$$- \frac{\sin \phi}{\sin \theta} P_n^1(\cos \theta) \frac{1}{kr} \frac{d}{dr} [r j_n(kr)] \hat{\phi}$$

# Holographic Tracking & Characterization

## How Lorenz-Mie Microscopy works

$$E_s(kr) = E_0 \sum_{n=1}^{\infty} i^n \frac{2n+1}{n(n+1)} \left( i \color{red} a_n \color{black} N_{e1n}^{(3)}(kr) - \color{red} b_n \color{black} M_{o1n}^{(3)}(kr) \right)$$

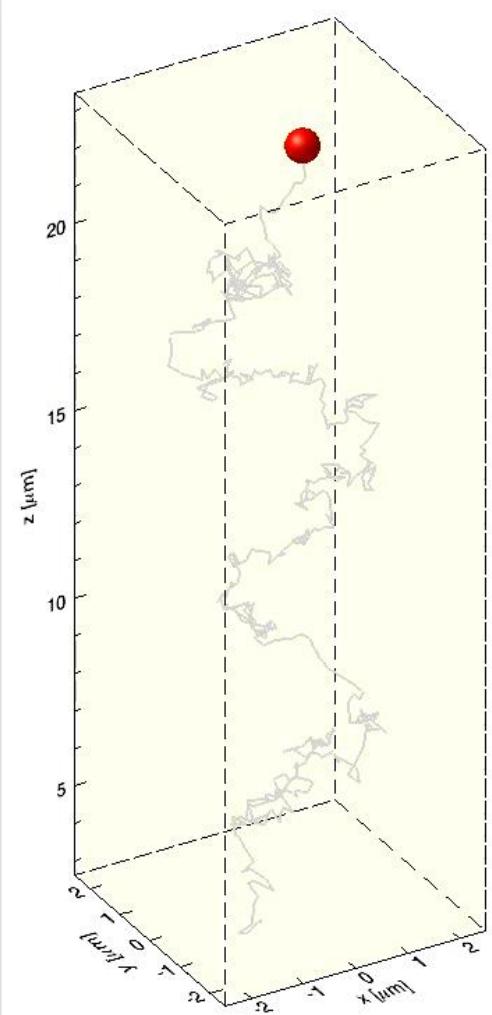
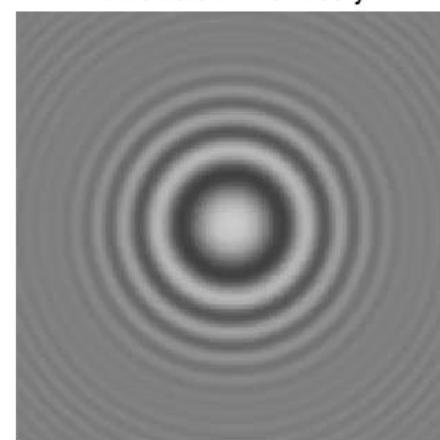
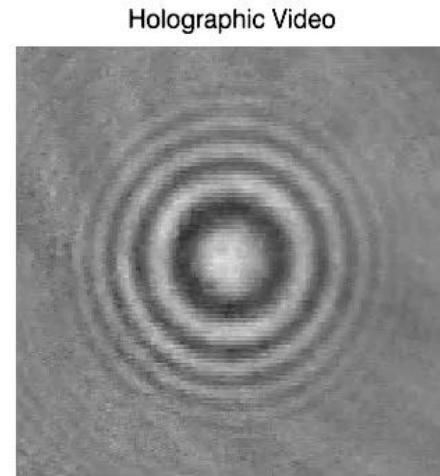
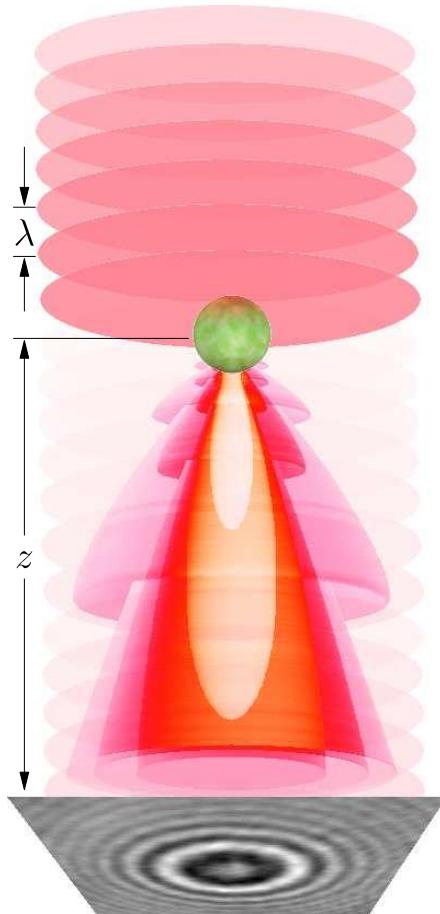
Lorenz-Mie scattering coefficients for a sphere:

$$\color{red} a_n \color{black} = \frac{m^2 j_n(mka_p) [ka_j_n(ka_p)]' - j_n(ka_p) [mka_p j_n(mka_p)]'}{m^2 j_n(mka_p) [ka_p h_n^{(1)}(ka_p)]' - h_n^{(1)}(ka_p) [mka_p j_n(mka_p)]'}$$

$$\color{red} b_n \color{black} = \frac{j_n(mka_p) [ka_p j_n(ka_p)]' - j_n(ka_p) [mka_p j_n(mka_p)]'}{j_n(mka_p) [ka_p h_n^{(1)}(ka_p)]' - h_n^{(1)}(ka_p) [mka_p j_n(mka_p)]'}$$

- relative radius:  $ka_p$
- refractive index:  $m = \frac{n_p}{n_m}$

# Lorenz-Mie Microscopy



Lee et al., *Optics Express* **15**, 18275 (2007)

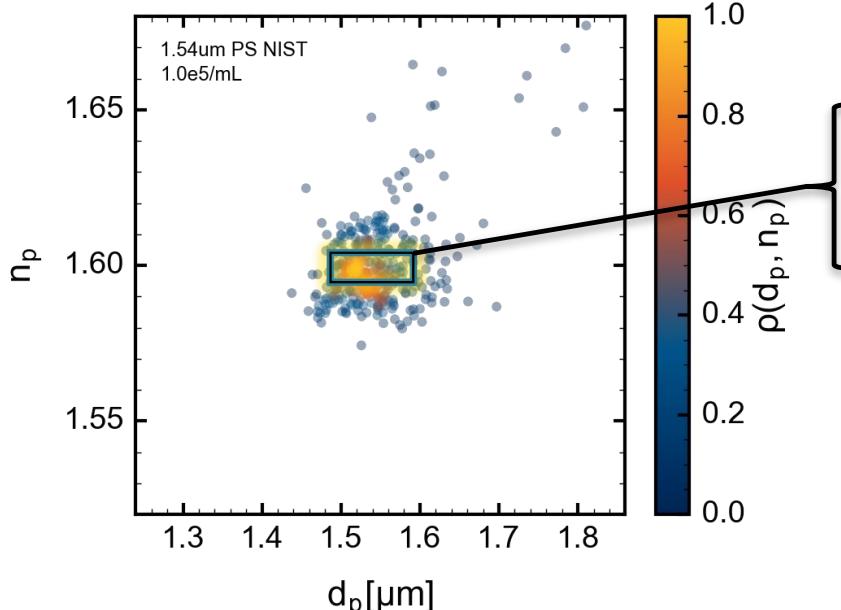
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# Holographic Tracking & Characterization

Independently verified performance

Property	Precision	Range
3D Position	$\Delta r_p < 3 \text{ nm}$	$100 \times 100 \times 100 \mu\text{m}^3$
Radius	$\Delta a_p \leq 2 \text{ nm}$	$200 \text{ nm} - 20 \mu\text{m}$
Refractive Index	$\Delta n_p \leq 10^{-3}$	$1 - 5$
Concentration	$\Delta c_p \approx \sqrt{c_p}$	$10^3 \text{ mL}^{-1} - 10^8 \text{ mL}^{-1}$



NIST Traceable Particles

$$d_p = 2 a_p = 1.54 \pm 0.05 \mu\text{m}$$

$$n_p = 1.598 \pm 0.005$$

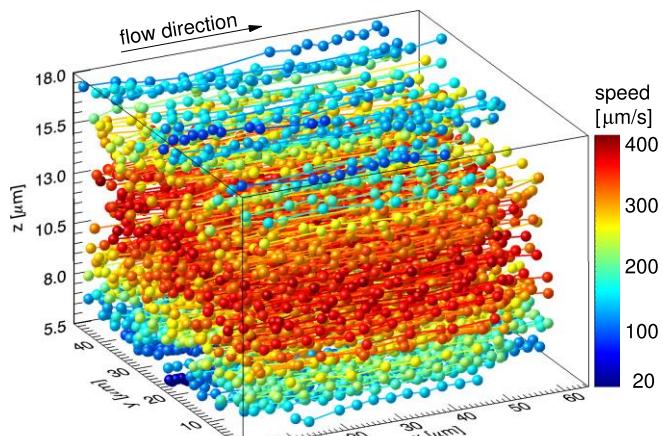
Bangs Labs #12035



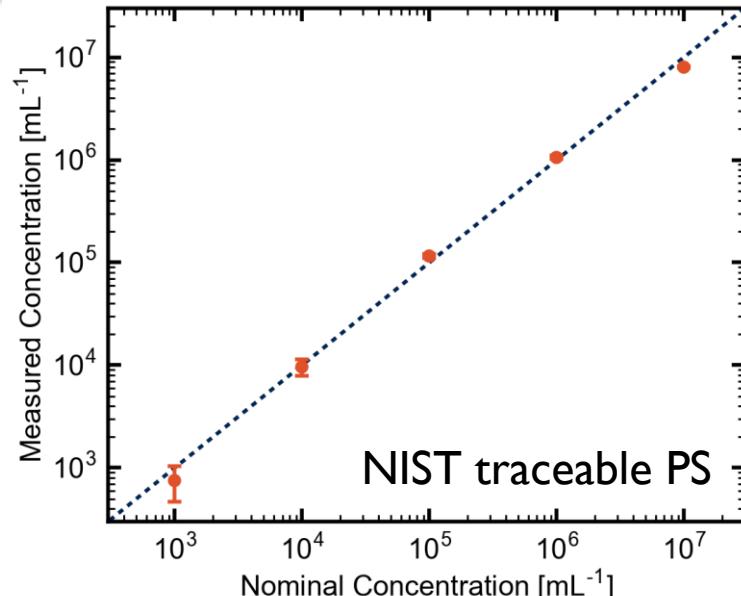
# Holographic Tracking & Characterization

Independently verified performance

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Radius	$\Delta a_p \leq 2 \text{ nm}$	$200 \text{ nm} - 20 \mu\text{m}$
Refractive Index	$\Delta n_p \leq 10^{-3}$	$1 - 5$
Concentration	$\Delta c_p \approx \sqrt{c_p}$	$10^3 \text{ mL}^{-1} - 10^7 \text{ mL}^{-1}$

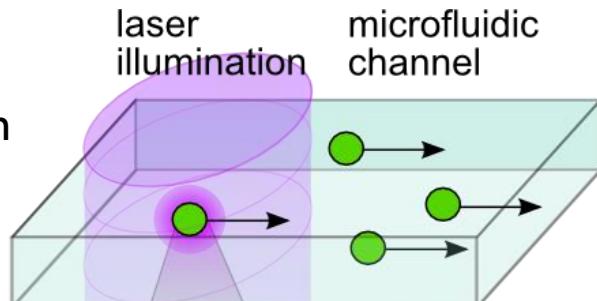


Measured flow volume  
(no calibration)

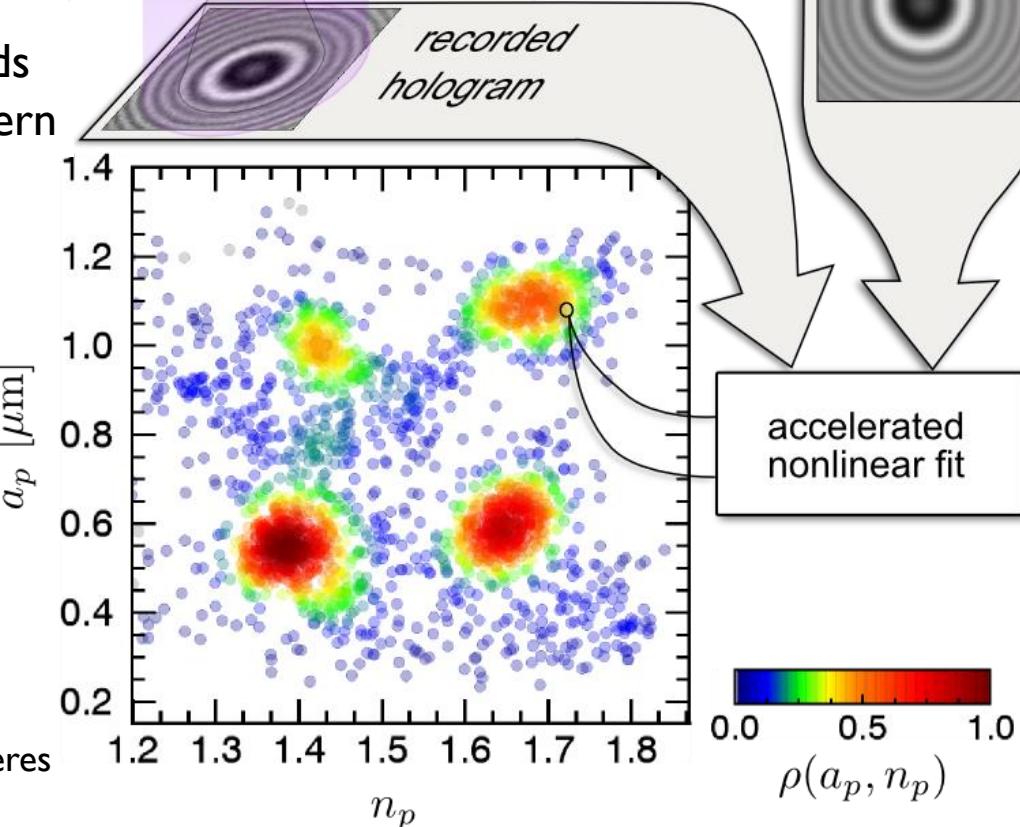


# Automated Holographic Characterization

1. Particles pass through laser beam in microfluidic channel

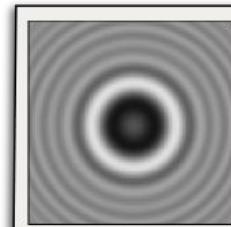


2. Microscope records interference pattern



Lorenz-Mie theory

$$|\hat{x} + e^{ikz_p} \mathbf{f}_s(k(\mathbf{r} - \mathbf{r}_p))|^2$$

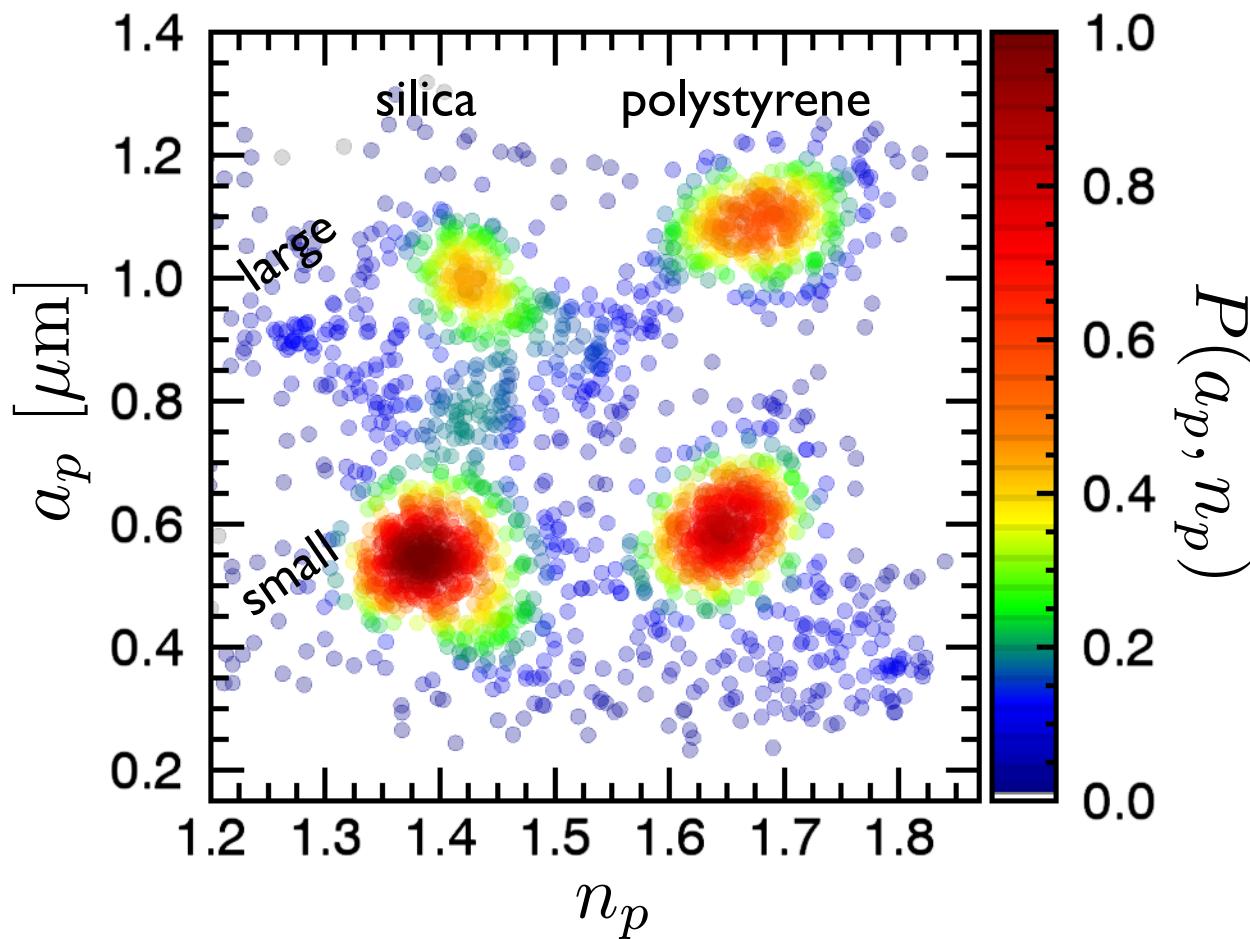


3. Compare measurement to scattering theory

5. Build statistics:  
Each point  
characterizes  
one particle

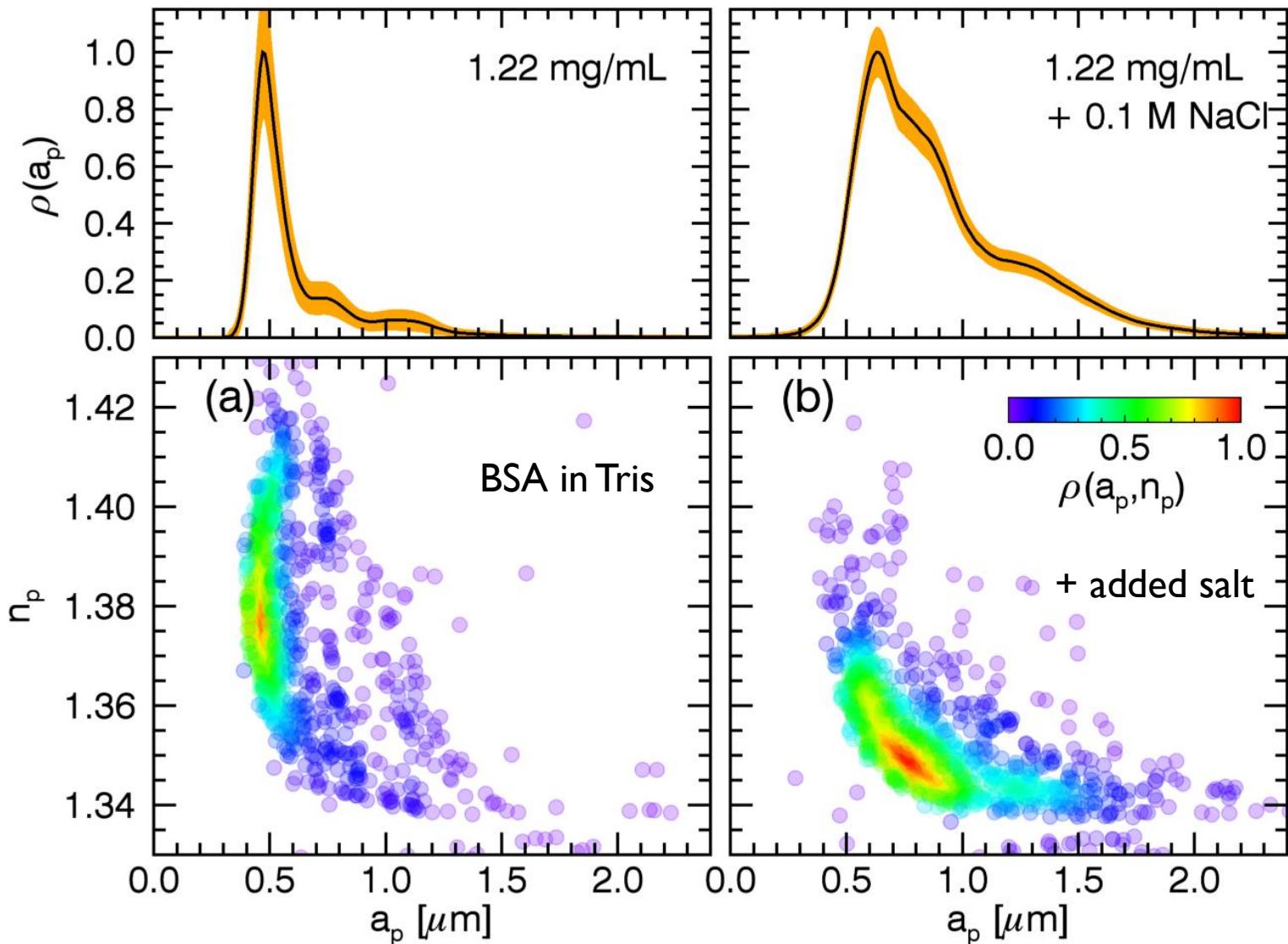
4. Comparison yields  
 $a_p$  : radius  
 $n_p$  : refractive index  
 $\mathbf{r}_p$  : 3D position

# Characterizing Colloidal Mixtures



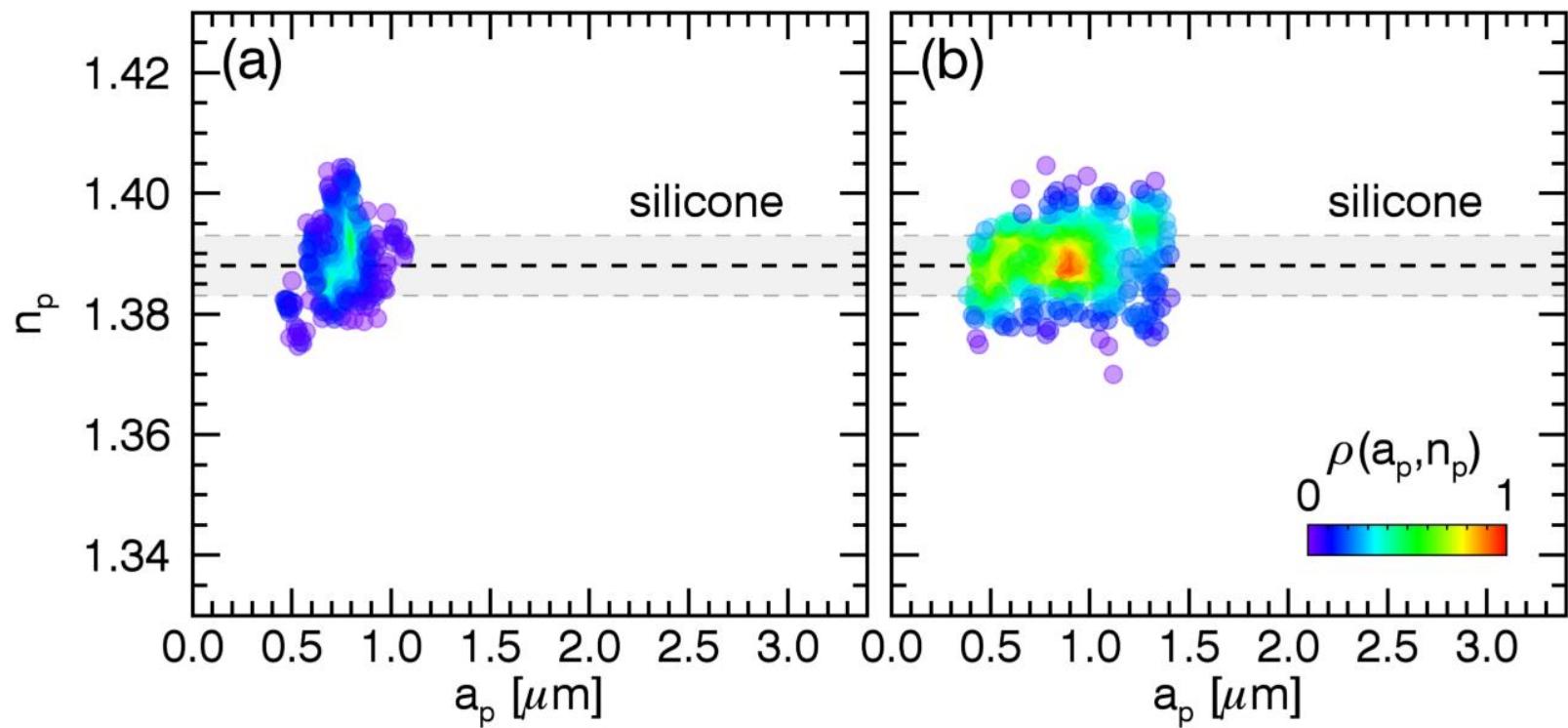
Yevick, Hannel & Grier, *Optics Express* **22**, 22864 (2014)

# Characterizing Protein Aggregates

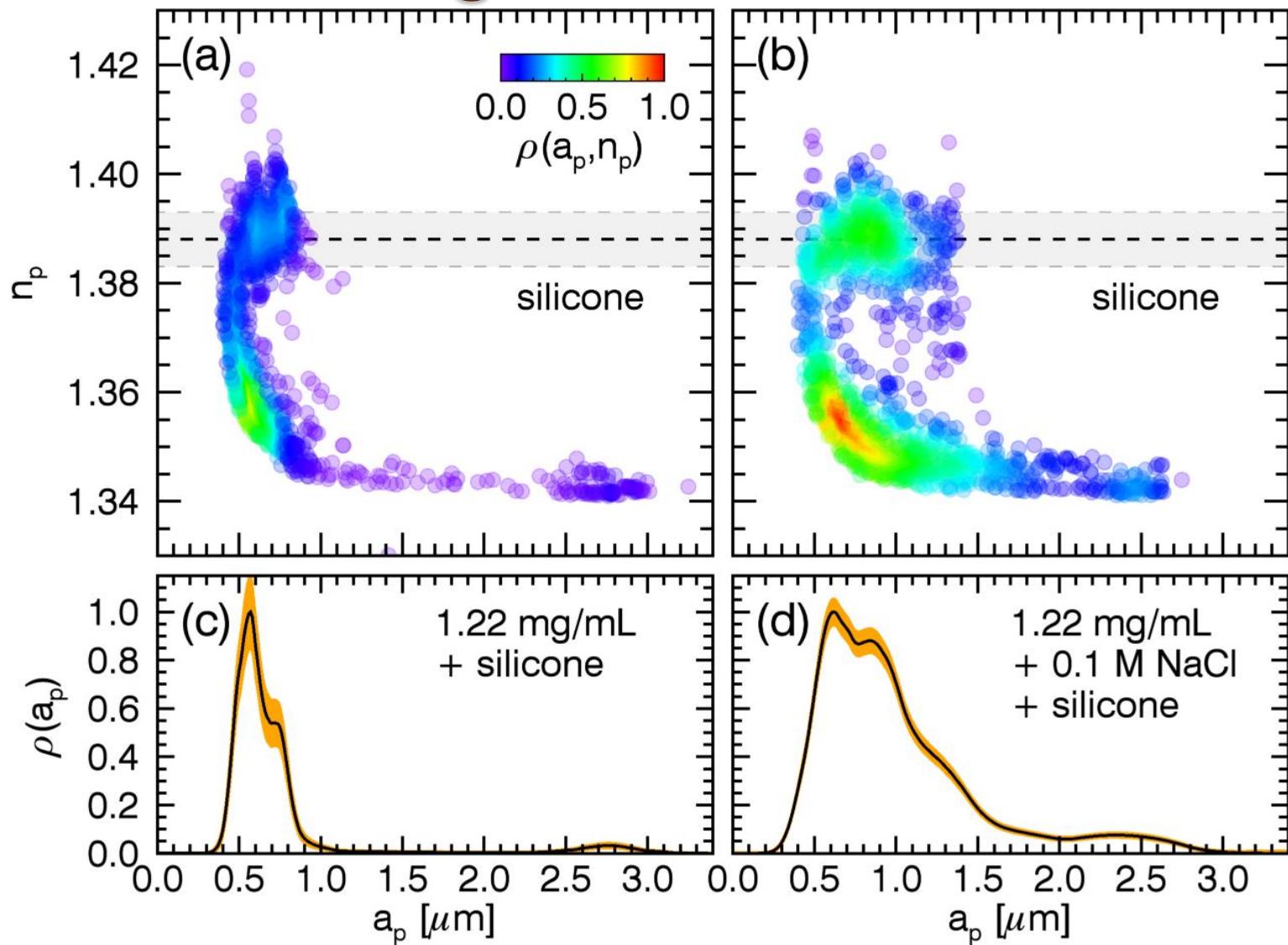


Wang et al., *J. Pharm. Sci.* **105**, 1074 (2016)

# Differentiating Silicone Oil Droplets



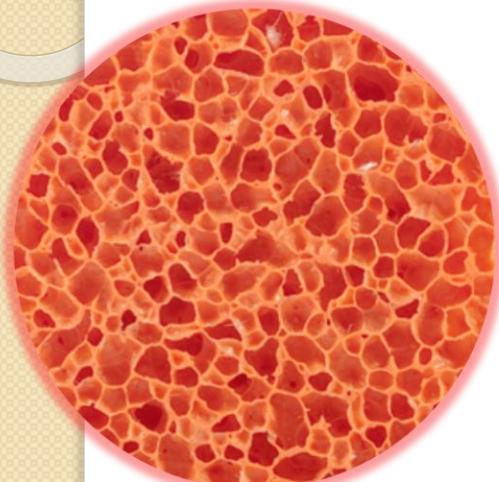
# Characterizing Protein & Contaminants



Wang et al., *J. Pharm. Sci.* **105**, 1074 (2016)

# Effective Medium Theory

Porous particle



Material: Refractive index:  $n$

Volume fraction:  $\phi$

Medium: Refractive index:  $n_m$

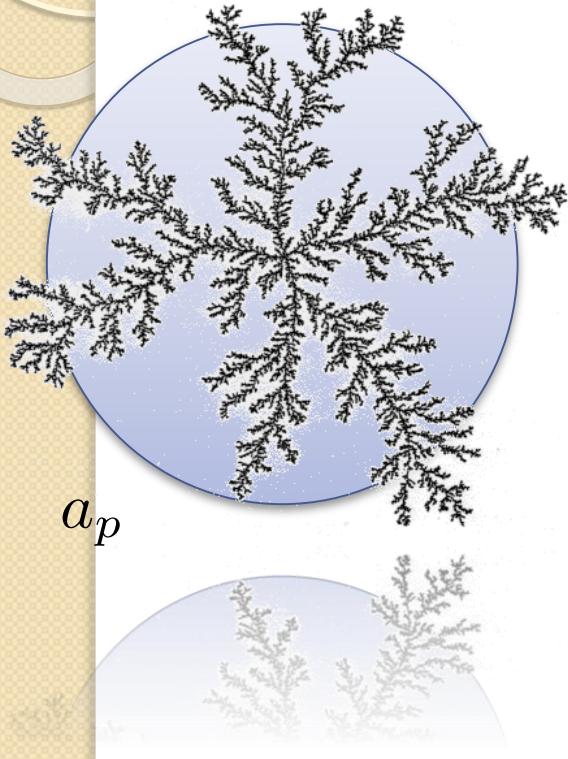
Effective refractive index:  $n_p$

$$f(n_p) = \phi f(n) + (1 - \phi)f(n_m)$$

Lorentz-Lorenz factor:

$$f(n) = \frac{n^2 - n_m^2}{n^2 + 2n_m^2}$$

# Fractal Aggregates



Volume fraction:

$$\phi = \left( \frac{a_0}{a_p} \right)^{3-D}$$

Effective Medium Theory:

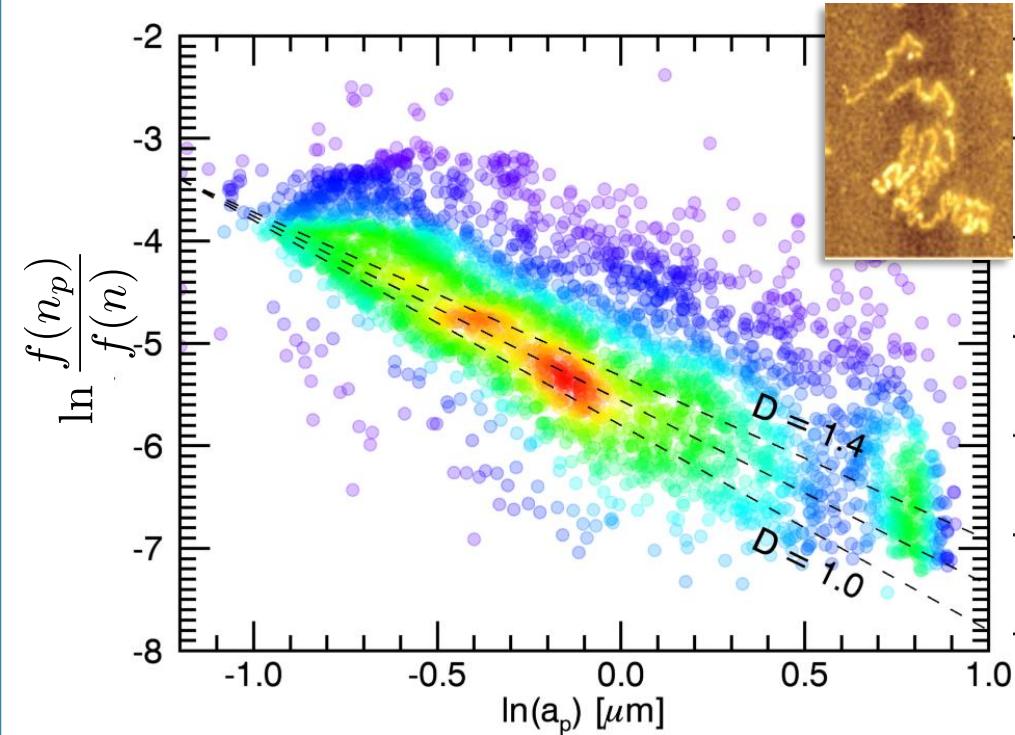
$$f(n_p) = \phi f(n) + (1 - \phi) f(n_m)$$

Size-Index Scaling:

$$\frac{f(\textcolor{red}{n}_p)}{f(n)} = \left( \frac{a_0}{\textcolor{red}{a}_p} \right)^{3-D}$$

# Fractal Scaling of Protein Aggregates

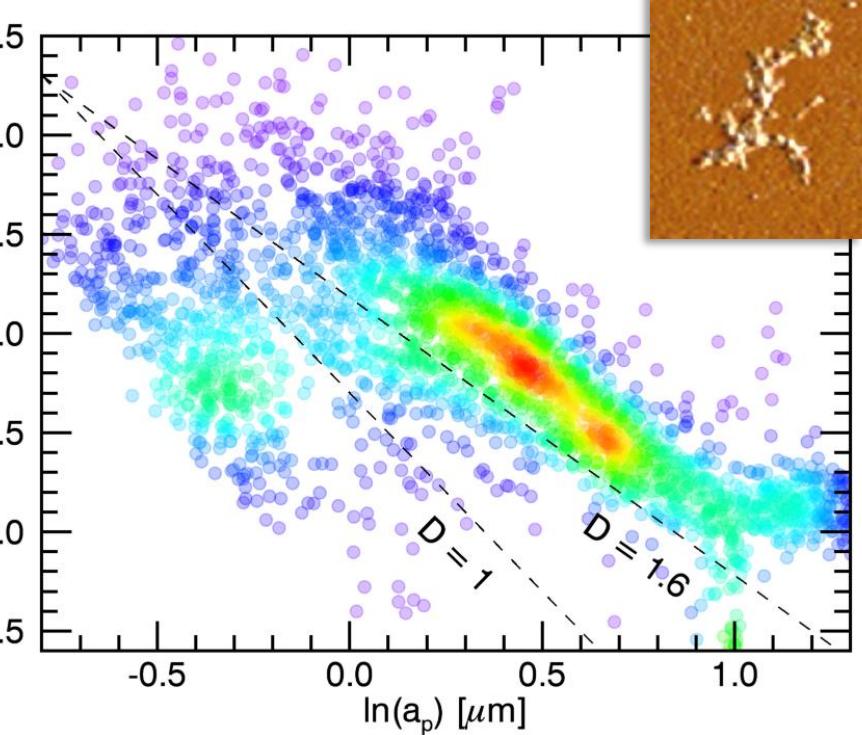
Bovine Serum Albumin



$$D = 1.1 \pm 0.1$$

Filamentary clusters

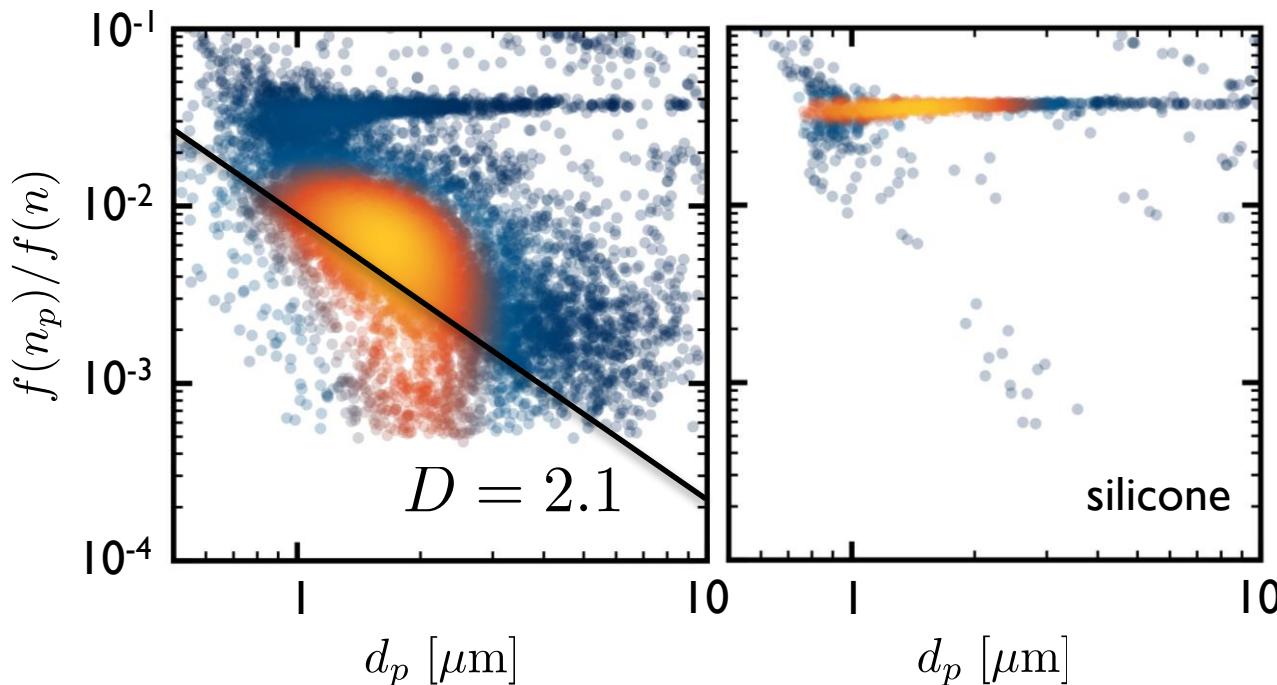
Bovine Insulin



$$D = 1.6 \pm 0.1$$

Cluster-cluster aggregates

# Holographic Characterization of Human IgG



Large fractal dimension: compact clusters

Differentiation between protein aggregates and silicone droplets



# Holographic Characterization

- Detects, counts and characterizes subvisible protein aggregates *in situ*
- Differentiates by composition & morphology
- Fast, time-resolved measurements
- Independently verified precision & accuracy
- Minimal calibration





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# Comparison with Industry Standard

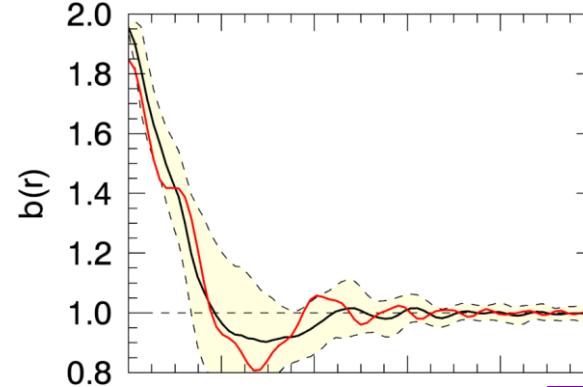
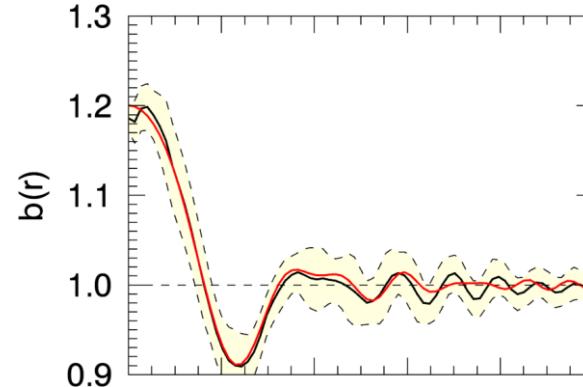
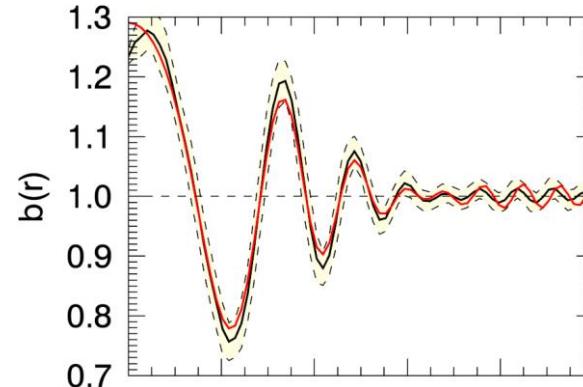
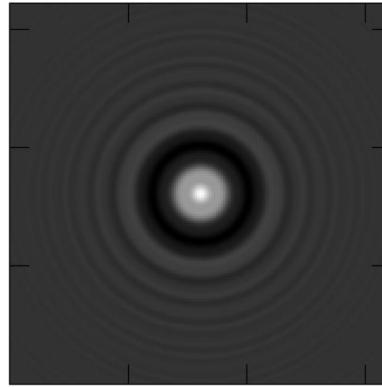
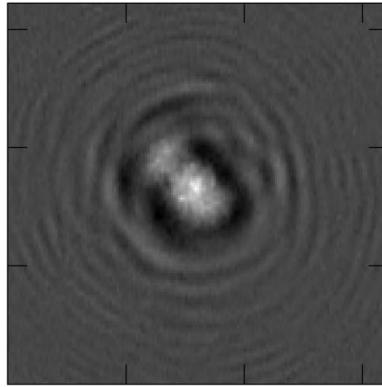
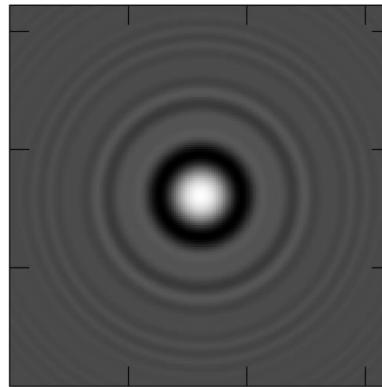
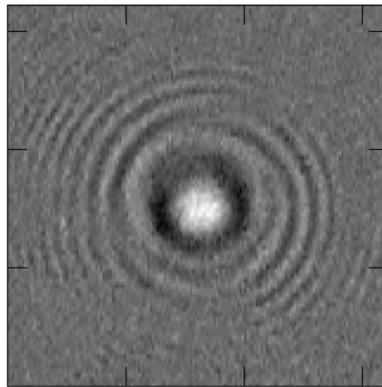
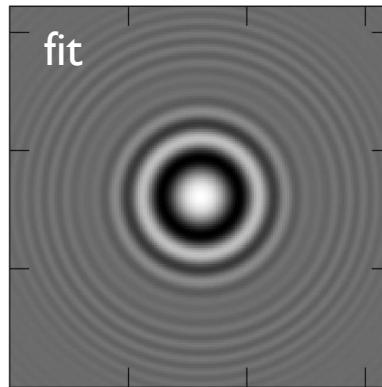
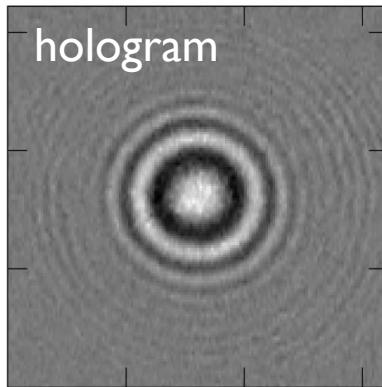
## DLS (Malvern Zetasizer Nano)

diameter [ $\mu\text{m}$ ]	manufacturer	Zetasizer [ $\mu\text{m}$ ]	stdev(ap)	HVM(ap) [ $\mu\text{m}$ ]	stdev(ap)	HVM(np) [ $\mu\text{m}$ ]	Porosity
15.17 $\pm$ 0.14	PS-research Particle	-	-	15.282	0.6	1.599 $\pm$ 0.02	1%
12	PS-research Particle	-	-	12.07	0.13	1.597 $\pm$ 0.004	1%
10.02 $\pm$ 0.08	PS-research Particle	-	-	9.928	1.1	1.605 $\pm$ 0.008	1%
1.51 $\pm$ 0.045	Polyscience	-	-	1.634	0.04	1.576 $\pm$ 0.008	12.10%
0.995 $\pm$ 0.021	Duke scientific	0.972	0.13	1.1	0.01	1.58 $\pm$ 0.005	6%
0.701 $\pm$ 0.006	Duke scientific	0.8166	0.113	0.824	0.04	1.565 $\pm$ 0.016	25%
0.304 $\pm$ 0.006	Duke scientific	0.37	0.088	0.482	0.03	x	x
0.06 $\pm$ 0.0025	Duke scientific	0.082	0.045	0.078	0.02	x	x

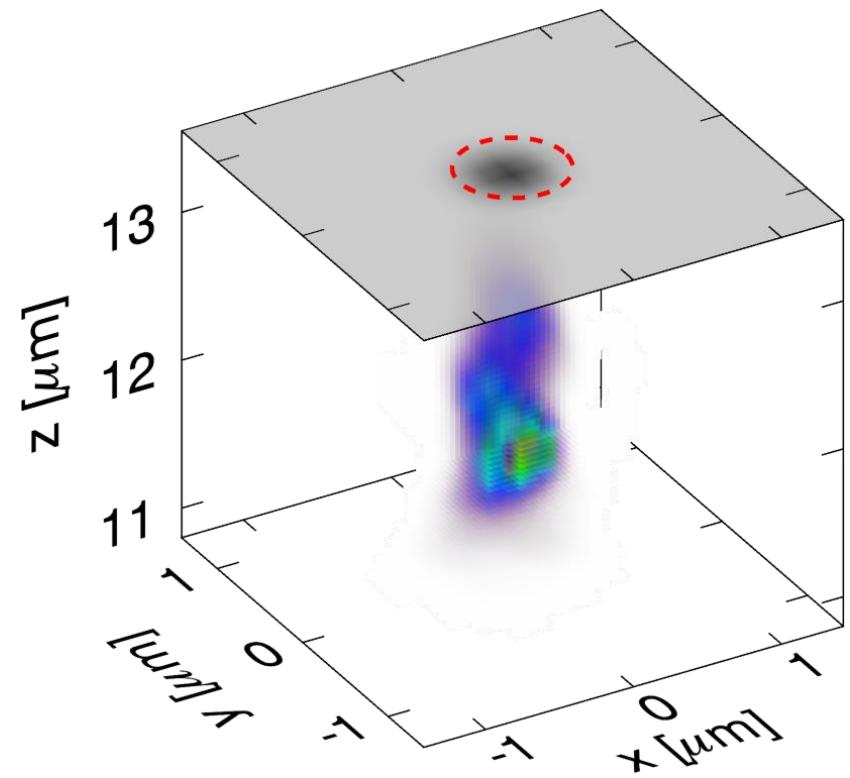
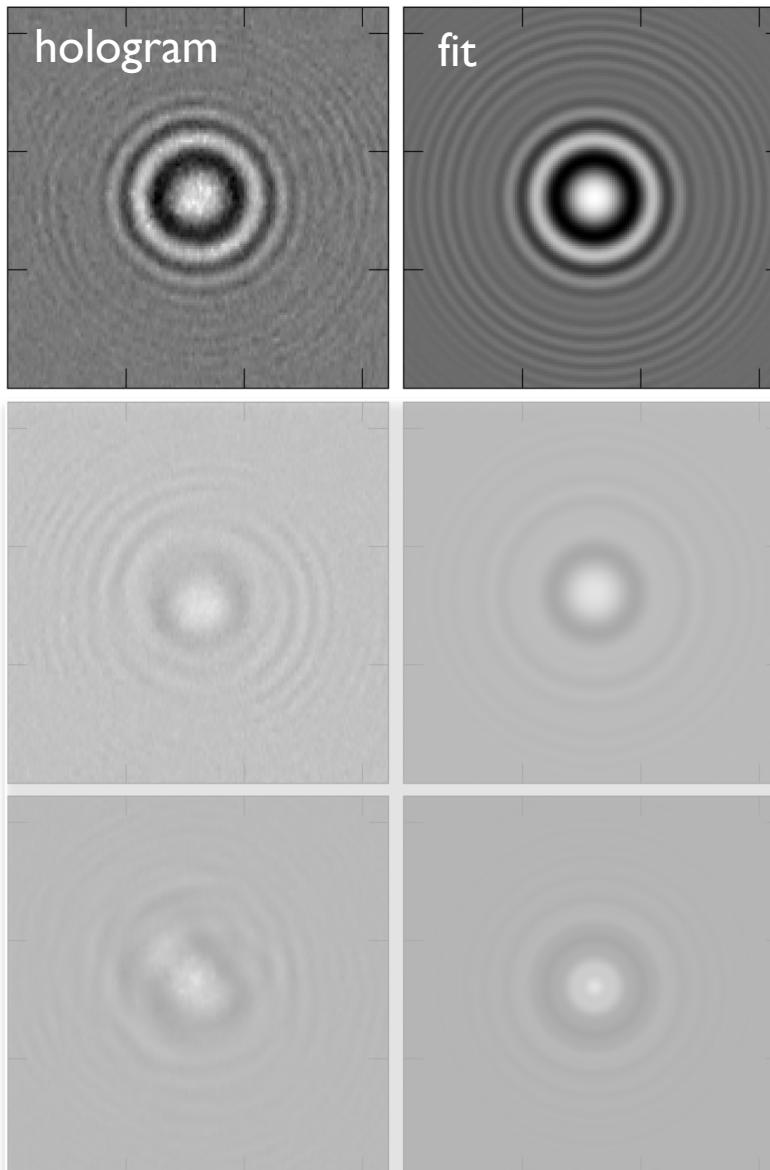
- 1. Consistent mean radius results in common range
- 2. HVM also works for larger particles
- 3. HVM results have smaller (better) instrumental spread
- 4. HVM also yields refractive index (porosity)

Cheong et al., *Opt. Express* **17**, 13071 (2009)  
 Cheong, Xiao, Pine & Grier, *Soft Matter* **7**, 6816 (2011)

# So, what do our clusters look like?



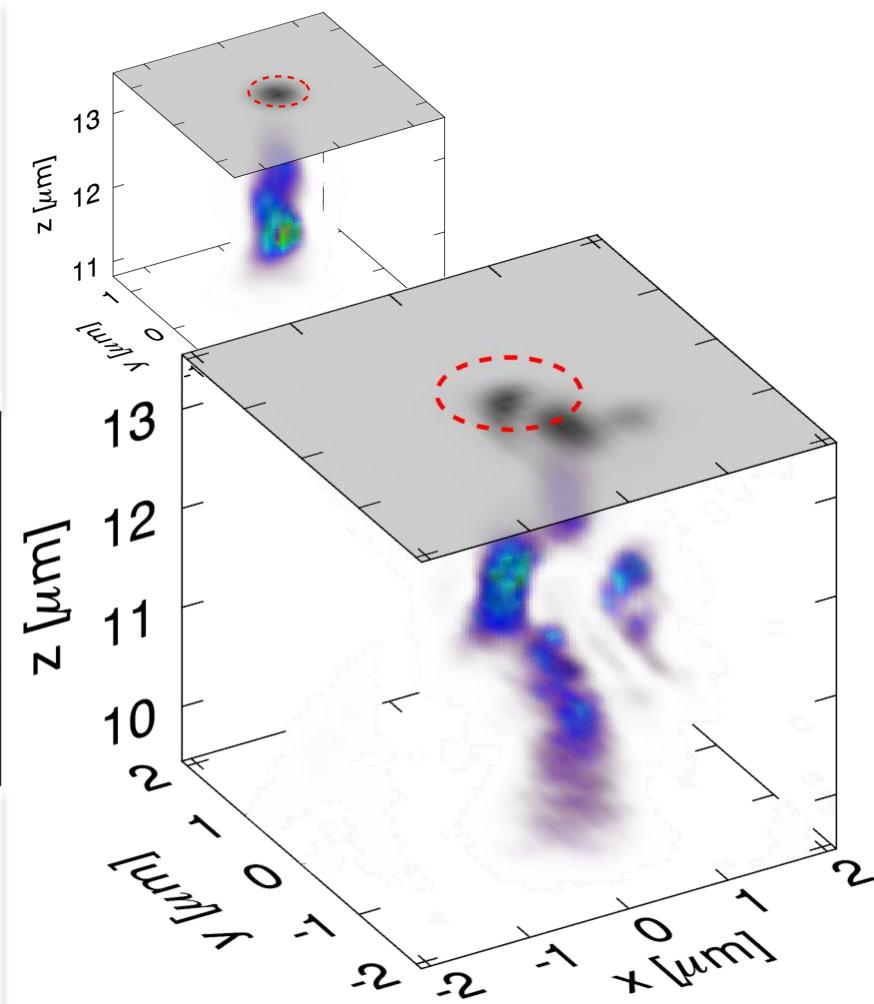
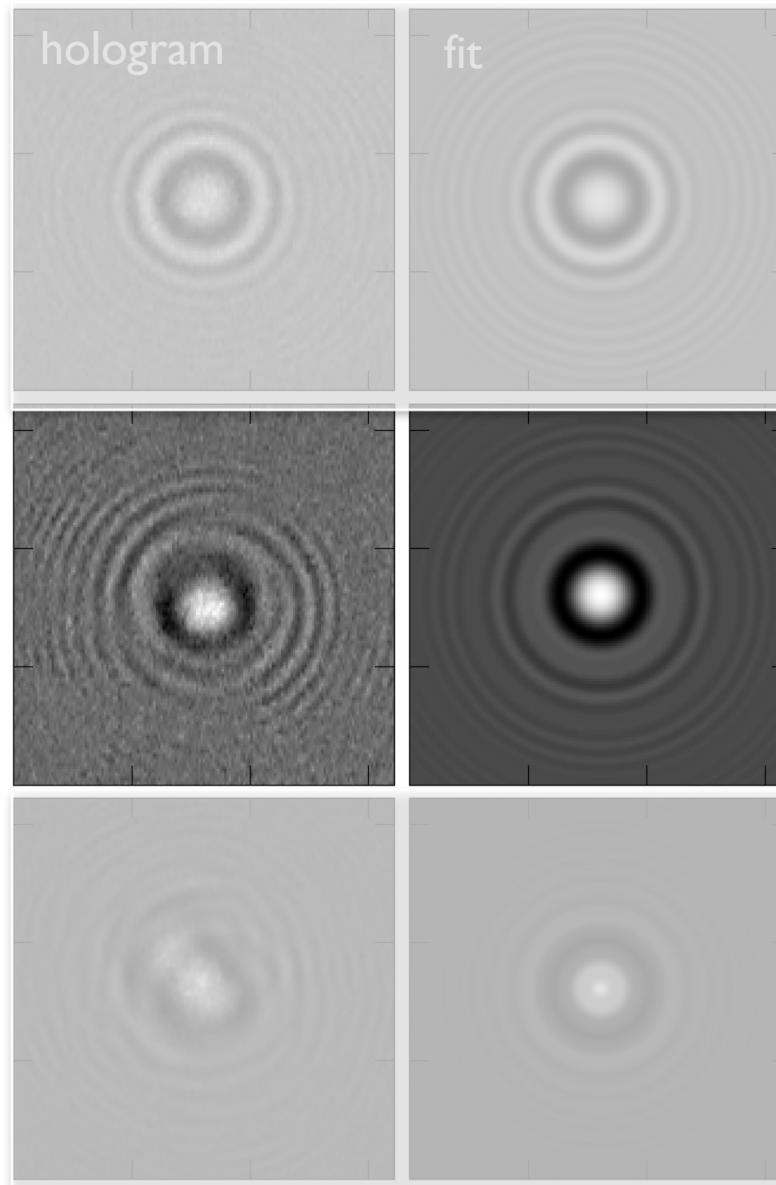
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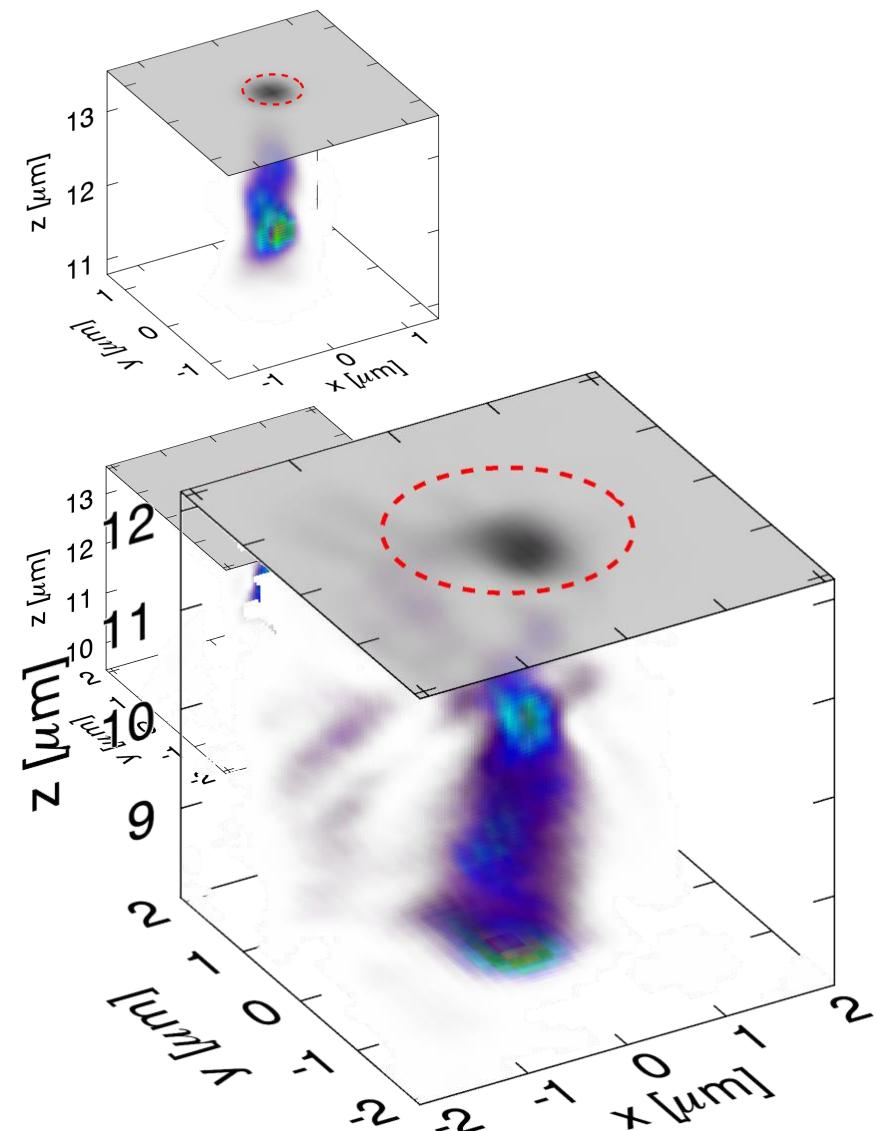
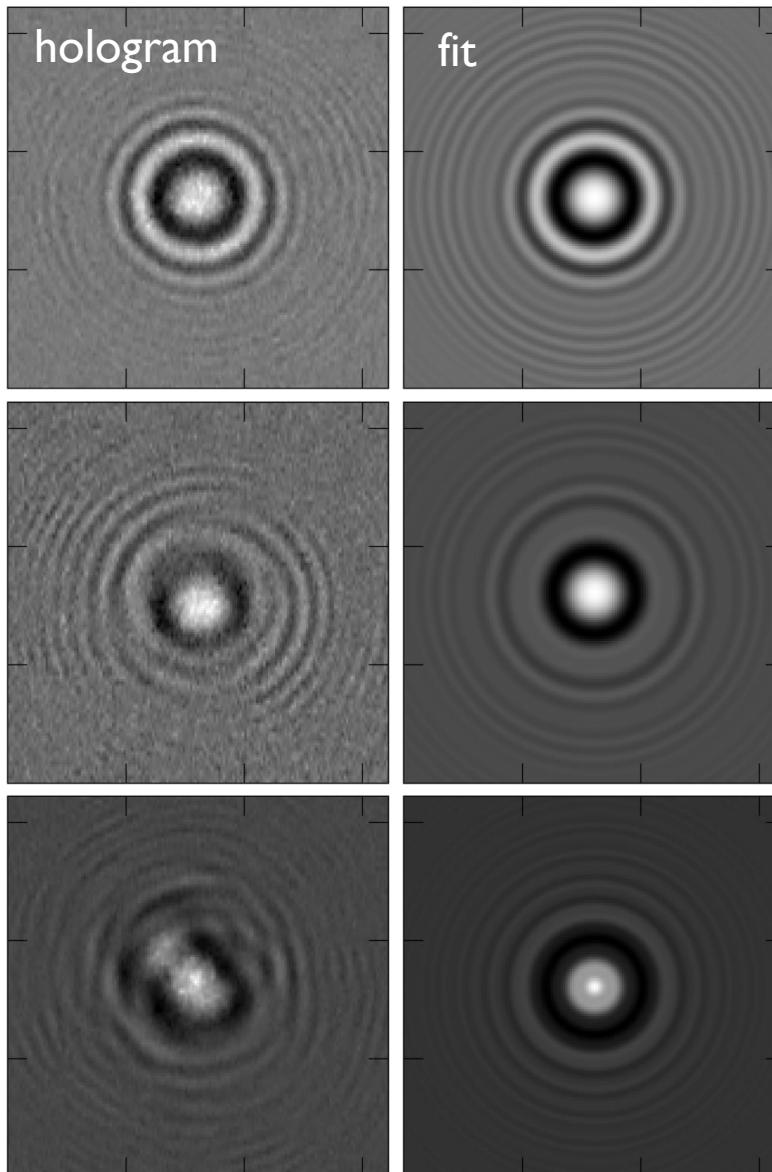
## Holographic Deconvolution Microscopy

Dixon *et al.*, *Opt. Express* **19**, 16410 (2011)  
Chen *et al.*, *J. Pharm. Sci.* **105**, 1074 (2016)

# So, what do our clusters look like?



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