

WATER NMR FOR PROTEIN AGGREGATION CHARACTERIZATION

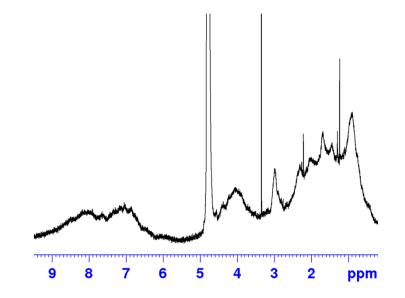
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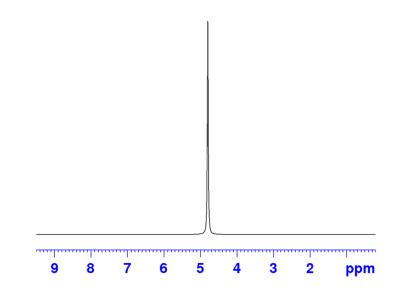
December 5, 2016, Baltimore **Protein Aggregation Measurement in Biotherapeutics: Established and Emerging Techniques**

WATER NMR-A NUISANCE OR A TOOL?

In aqueous solutions, solute resonances are considered the most important for NMR, and deuteration or suppression is used to remove interfering water signal



BSA (15 mg/mL in PBS buffer) with water suppression (100 scans): bad resolution of protein resonances.



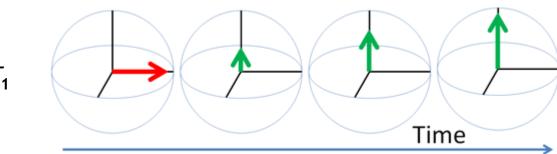
BSA (15 mg/mL in PBS buffer) without water suppression (1 scan): high S/N for narrow water signal, protein resonances are invisible.



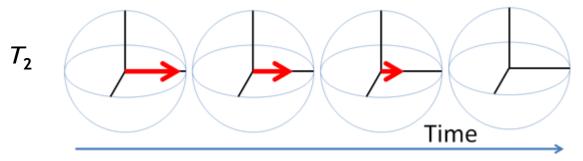
Y. Feng, M. Taraban & Y.B. Yu (2015) Chem. Commun. 51, 6804

RELAXATION RATES IN NMR

 T_1



 T_1 is spin-lattice or longitudinal relaxation, equal to time of energy transfer from excited to ground state along *z*-axis, often is defined by interaction between nucleus and media (solvent, diffusion).

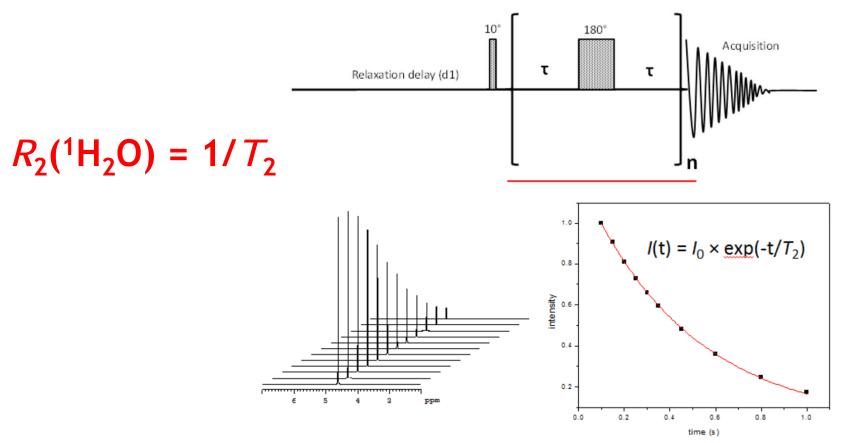


 T_2 is spin-spin or transverse relaxation, equal to time of energy transfer within the nucleus in the *xy*-plane due to dephasing, NMR line broadening down to the disappearance of the NMR signal, often defined by dipolar interactions, anisotropy of molecule, etc.

Relaxation of the nucleus to its ground state (aligned with external magnetic field) is controlled by two mechanisms.



TRANSVERSE RELAXATION OF WATER CPMG PULSE SEQUENCE

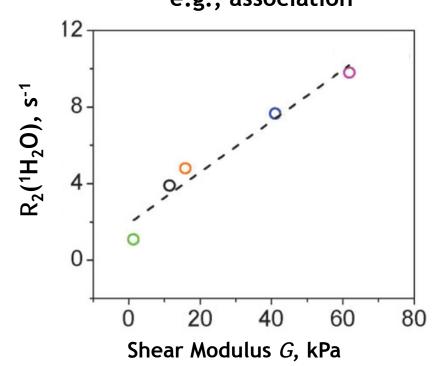


 T_2 is measured using classic CPMG pulse sequence that allows to monitor the drop in magnetization in *xy*-plane.

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WATER NMR-A TOOL

Water signal carries information on the global changes in the solute-water molecules interact with solute molecules and become sensitive to its changes, e.g., association

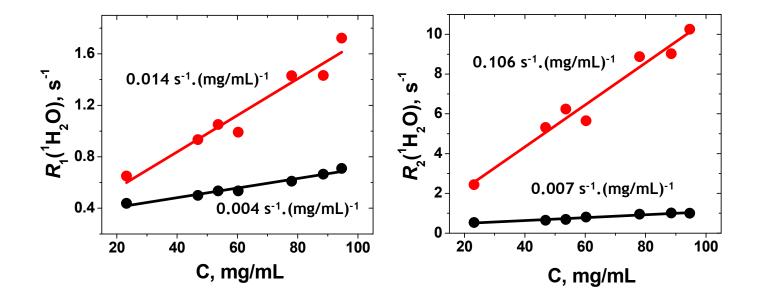


Water proton transverse relaxation rate, $R_2({}^{1}H_2O)$, could be used to measure the stiffness of peptide-based hydrogel. Gelation and aggregation both involve association, so would $R_2({}^{1}H_2O)$ also be sensitive to protein aggregation?

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PRIOR ART—WATER RELAXATION IN HEAT-DENATURED PROTEINS

Ovalbumin • fresh; • heat denatured (0.33 Tesla, 14 MHz ¹H)



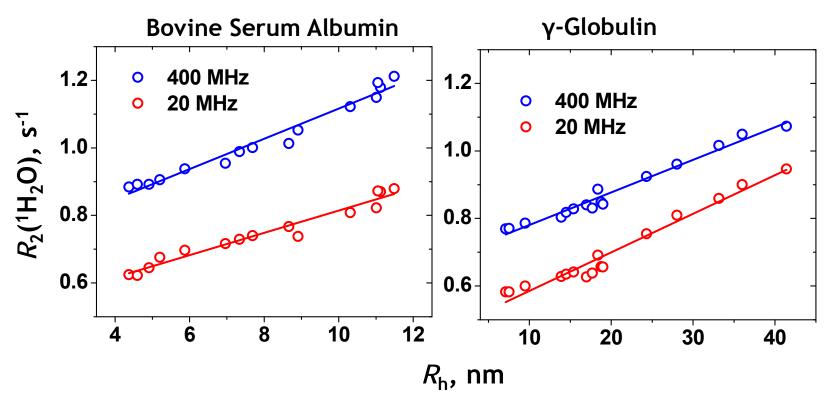
Does $R_2({}^{1}H_2O)$ correlate with aggregate size?



Daszkiewicz et al. (1963) Nature, 200, 1006; Oakes et al. (1976) J. Chem. Soc. Faraday Trans. I 72, 228; Hills et al. (1989) Mol. Phys. 67, 903; Indrawati et al. (2007) J. Sci. Agric. 87, 2207

A PROBE FOR PROTEIN AGGREGATION

Water proton NMR is sensitive towards heat-induced aggregation of BSA and human γ -globulin, and could be used to quantify protein aggregation



 $R_2({}^{1}H_2O)$ linearly increases with the growth of average hydrodynamic radius of protein aggregates.

Similar sensitivity observed in high (400 MHz) and low-field (20 MHz, BT NMR)



GENERATION OF MONOCLONAL ANTIBODY AGGREGATES OF VARIOUS SIZES NONFILTERED mAb has been stressed by: **Jum FILTERED**

Freeze-Thaw (-40 °C \leftrightarrow 5 °C, 16 cycles) Heating at 50°C (36 h) Agitation (24 h)

Aggregation was studied by

Conventional Techniques Size-Exclusion Chromatography (SEC) Dynamic Light Scattering (DLS) Micro-Flow Imaging (MFI)

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Water NMR (W/NMR) Transverse Relaxation Rate of Water $R_{2}(^{1}H_{2}O)$

UNSTRESSED CONTROL

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STRESS

S

0.45 µm FILTERED

COMPARE

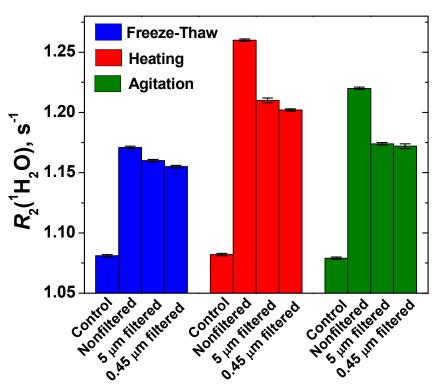
- Aggregates ≥ 5 µm
- Aggregates 0.45 to 5 µm
- Aggregates ≤ 0.45 µm





MEASUREMENT OF MAB AGGREGATION BY WNMR

 $R_2({}^{1}H_2O)$ responded to aggregate formation under different stresses and differs from control after filtration



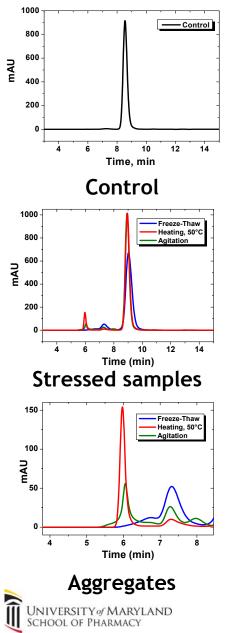
- R₂(¹H₂O) increased in each stressed sample compared to the unstressed control sample
- Filtration reduced the increase in $R_2({}^{1}H_2O)$ for all stresses
- $R_2({}^{1}\text{H}_2\text{O})$ was still different after 0.45 micron filtration between stresses

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MEASUREMENT OF MAB AGGREGATION BY SEC

MedImmune



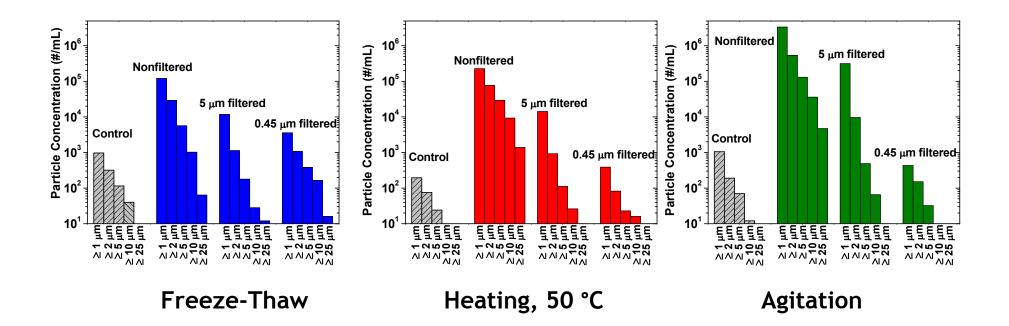
% Low (LMW) and % High Molecular Weight (HMW) and Total % soluble mAb aggregates for three stresses

	Aggregates	Control	Nonfiltered	5 μm filtered	0.45 μm filtered
Freeze- Thaw	% LMW	0.8	7.4	7.3	7.3
	% HMW	0.0	1.9	2.0	2.0
	Total % Aggr	0.8	9.3	9.3	9.3
Heating 50°C	% LMW	0.8	1.4	1.4	1.4
	% HMW	0.0	6.5	6.4	6.4
	Total % Aggr	0.8	7.9	7.8	7.8
Agitation	% LMW	0.6	3.2	3.2	3.4
	% HMW	0.0	6.9	6.0	5.9
	Total % Aggr	0.6	10.1	9.2	9.3

- Total percentage of aggregates were similar, but aggregate profile was different between each stress type
- 5 μ m & 0.45 μ m filtration did not change the ratio between LMW and HMW aggregates or total percentage of aggregates

MEASUREMENT OF MAB AGGREGATION BY MFI

Decrease in particle counts during filtration from MFI



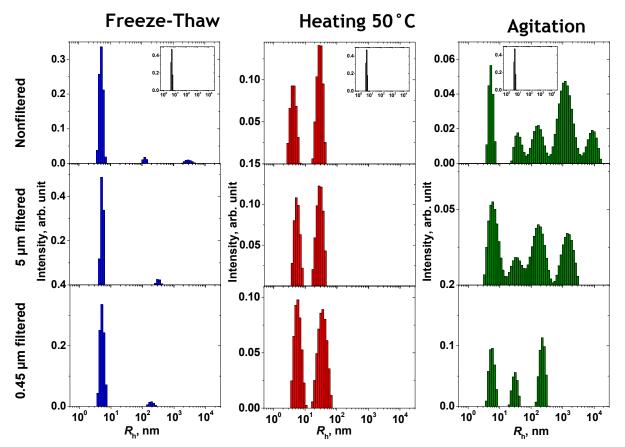
- 5 μ m filtration reduced particle counts from \geq 1 μ m to \geq 25 μ m (not only for \geq 5 μ m particles)
- After 0.45 μm filter, the samples for all three stresses are very close to the unstressed control





MEASUREMENT OF MAB AGGREGATION BY DLS

Each stress produced different particle size distributions (PSD) of aggregates

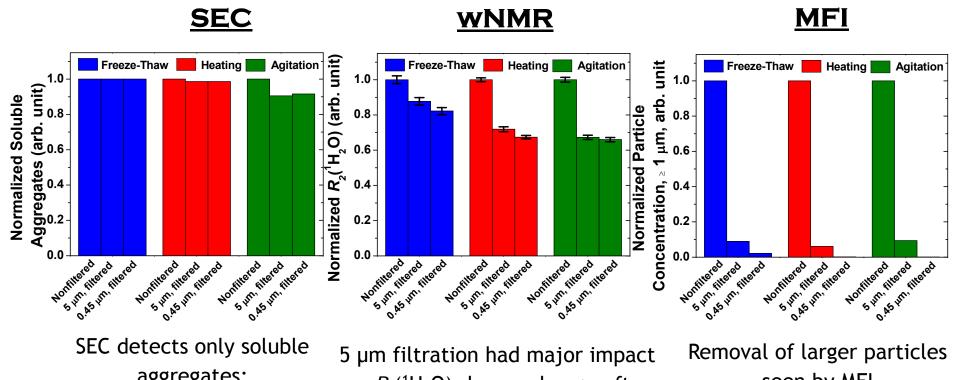


- 5 μ m and 0.45 μ m filters had minimal effect on PSD for freeze-thaw and heating
- Both filtration steps affected the PSD for agitation stress significantly



MedImmune

SENSITIVITY OF EACH TECHNIQUE TO SIZE AND NUMBER OF AGGREGATES



aggregates: Minimal sensitivity to sample filtration

on $R_2(^{1}H_2O)$, lesser change after 0.45 µm filtration, but differences still seen after 0.45 um filtration

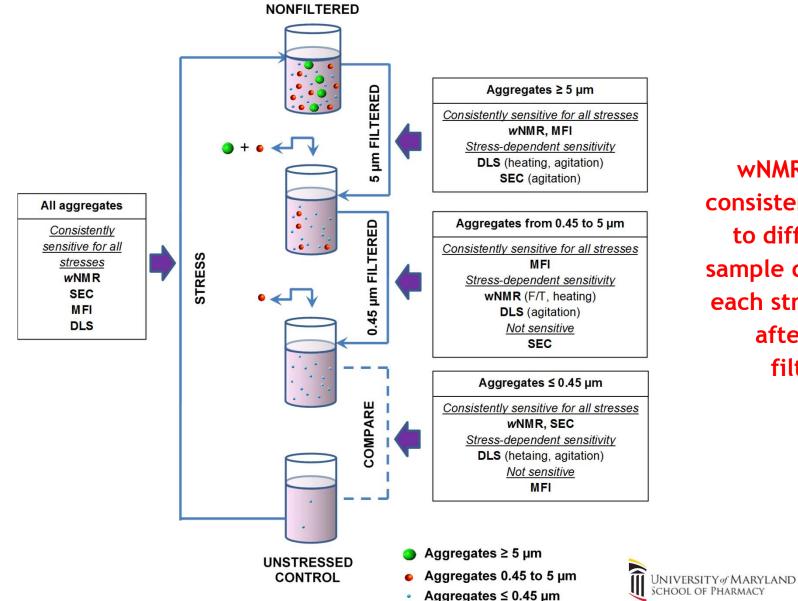
seen by MFI





Data are normalized by the difference between stressed sample and control so as fully stressed sample corresponds to 1, and control corresponds to 0.

SENSITIVITY RANGE OF EACH METHOD TO ANTIBODY AGGREGATES



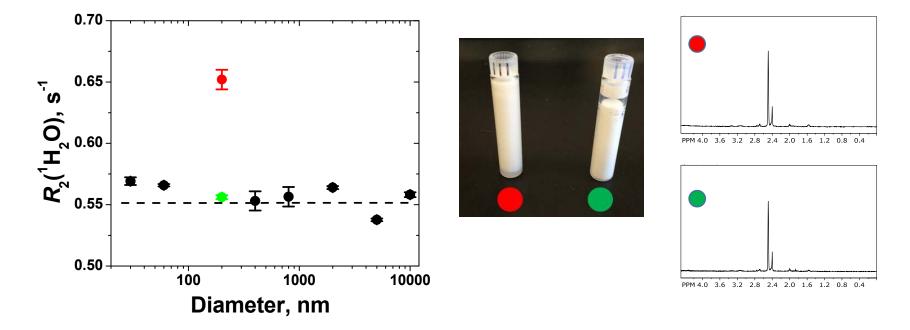
wNMR was most consistently sensitive to differences in sample quality across each stress type and after sample filtrations



POSSIBLE MECHANISMS OF SENSITIVITY OF PROTON NMR TO PROTEIN ÅGGREGATES

WATER NMR SENSES NANOPARTICLE CLUSTERING

- Two 200 nm polystyrene nanoparticle samples are visually indistinguishable
- ¹H NMR spectra show no difference in the signal intensities or chemical shifts



But one of them demonstrate anomalously high water relaxation rate

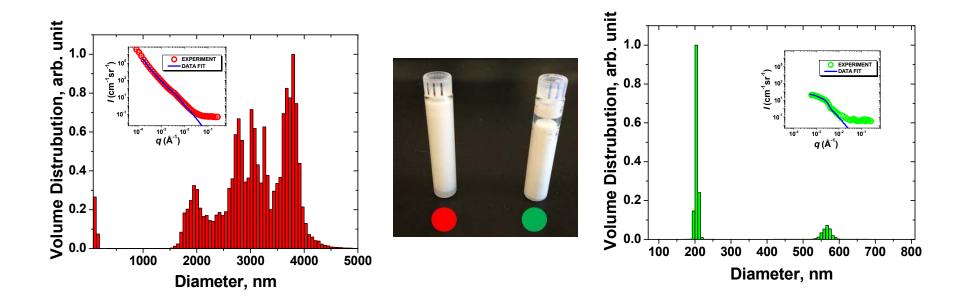
WHY?





WATER NMR SENSES NANOPARTICLE CLUSTERING

USAXS shows that anomalous sample contain mainly 2-4 µm particulates While the good quality sample overwhelmingly contains 200 nm particles



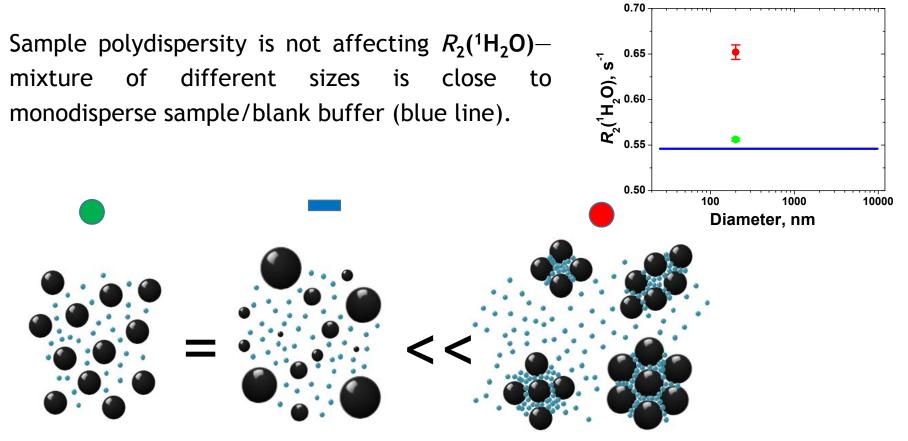
The anomalous sample had quality issues (confirmed by manufacturer), and nanoparticles in this sample are clustered and formed larger assemblies

USAXS = Ultra-small angle X-ray Scattering





WATER NMR SENSES NANOPARTICLE CLUSTERING



Water molecules in the clustered compartments have different diffusive exchange and local magnetic field gradient resulting in anomalously high $R_2({}^{1}\mathrm{H}_2\mathrm{O})$





CONCLUSIONS

- Water transverse relaxation rate $R_2({}^{1}H_2O)$ was a sensitive probe responding to changes in solute molecules: association, clustering, aggregation, etc.
- In protein aggregation, R₂(¹H₂O) was sensitive to the presence of insoluble particulates ≥ 5 µm, from ≥ 1 µm to 5 µm as well as to soluble protein aggregates below 1 um.
- $R_2({}^{1}H_2O)$ can be monitored noninvasively using inexpensive benchtop low-field NMR spectrometers with wide bore capable to accommodate drug product vials without opening or sampling.







DON'T THROW THE BABY OUT WITH THE BATHWATER!





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MedImmune

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Light Scattering Center University of Maryland

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