Towards Improved Characterization of MSCs

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MSC Ma	nufacturing
	Goal: 8 donors • Expand to ~ 100 T175 flasks • Change media every third day • Expand to ~ 80% confluence • Harvest at p3, p5, p7 • Cryopreserve, snap freeze, or initiate analytical procedures Yield ~ 2-3 x 10 ⁸ cells per passag University of the set of t



Publicly available sources Single method, single lab, uniform media One lot of FCS										
Sufficient MSCs for multiple characterization approaches Cell Manufacturing Information										
Designation*	Sex*	Age*	P3	P4	P5	P7	Cryopreserved	Comments		
PCBM 1641	F	23	124	169	116	93	502 x 10e6	P3, P5, P7		
PCBM1632	M	24	178		171	48	397 x 10e6	P3, P5, P7		
PCBM 1662	F	31	244		156	199	599 x 10e6	P3, P5, P7		
110877	M	22	194		251	106	551 x 10e6	P3, P5, P7		
8F3560	F	24	195		169	183	547 x 10e6	P3, P5, P7		
167696	F	22	208		121	166	495 x 10e6	P3, P5, P7		
127756	M	43	87		49		136 x 10e6	P3, P5		
PCBM 1655	F	47	35				35 x 10e6	P3		
	* Information	on supplied b	oy cell line s	ource						
								8		































Significance for Cell Therapy

- Quantitative functional assays; a crucial part of a systematic approach to identify and qualify predictive product characteristics for cell therapy products
 Consensus MSC Markers do not Correlate with Functional Heterogeneity: Donor or Tissue Culture Age Differences
 Robust enumeration of functional sub populations illustrates importance of extensive single cell analyses
- Assess
 - Differences between MSC donors
 - Impact of tissue culture conditions and duration
 - Correlation with other characteristics of MSCs
 - Enrichment techniques
- Application to understanding mechanisms controlling stem-cell differentiation and function

 - Heterogeneity
 Improved understanding of "stemness" Mechanisms of function





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