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The Nuts and Bolts of AntibioGrams in Long-Term Care Facilities

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Disclosures

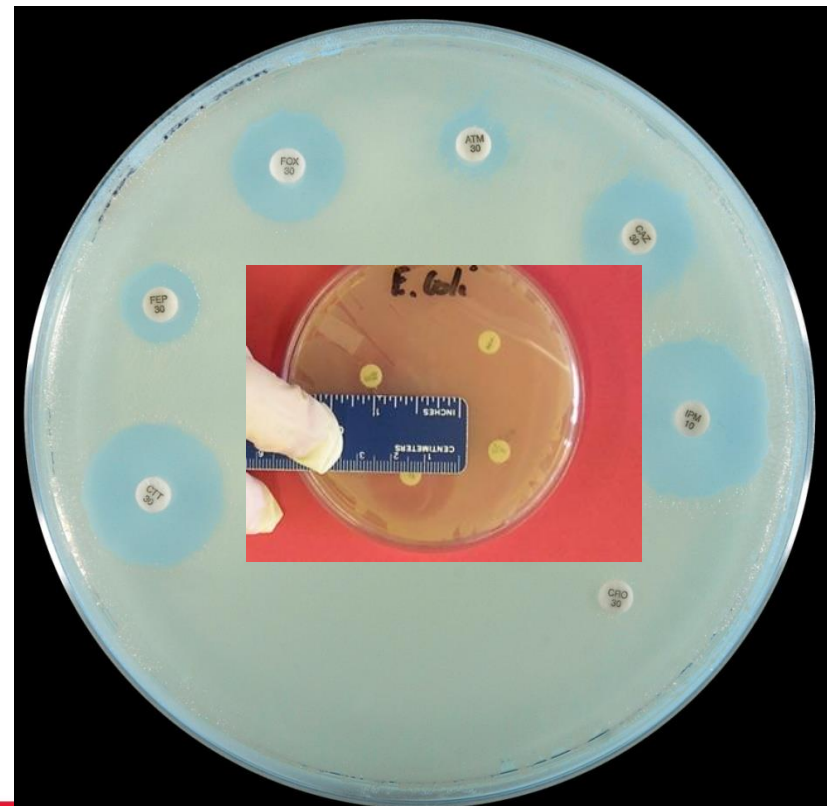
- Applied BioCode -Research Grant
- Beckman Coulter-Speaker
- bioMérieux-Speaker
- M39 Working group member

Objectives

- Review core concepts of antibiotic susceptibility and cumulative antibiotic susceptibility data/antibiograms
- Identify best practices for developing and maintaining annual antibiograms in long-term care
- Define how cumulative susceptibility data/antibiograms can be used in surveillance programs.

Antimicrobial Susceptibility Test

- Only performed on bacteria in which susceptibility to standardized treatment is not predictable.
 - Predictable
 - *β-Streptococcus*
 - Not-predictable
 - *E. coli*
- Antibiotics reported
 - Cascading antibiotics
 - Additional antibiotics from MDROs
 - What methods
 - Breakpoints used



Antibiotic Susceptibility Testing

Disk Diffusion

Kirby
Bauer

Qualitative

Dilution

Tube
Dilution

Agar
Dilution

Quantitative

Dilution and
Diffusion

E test

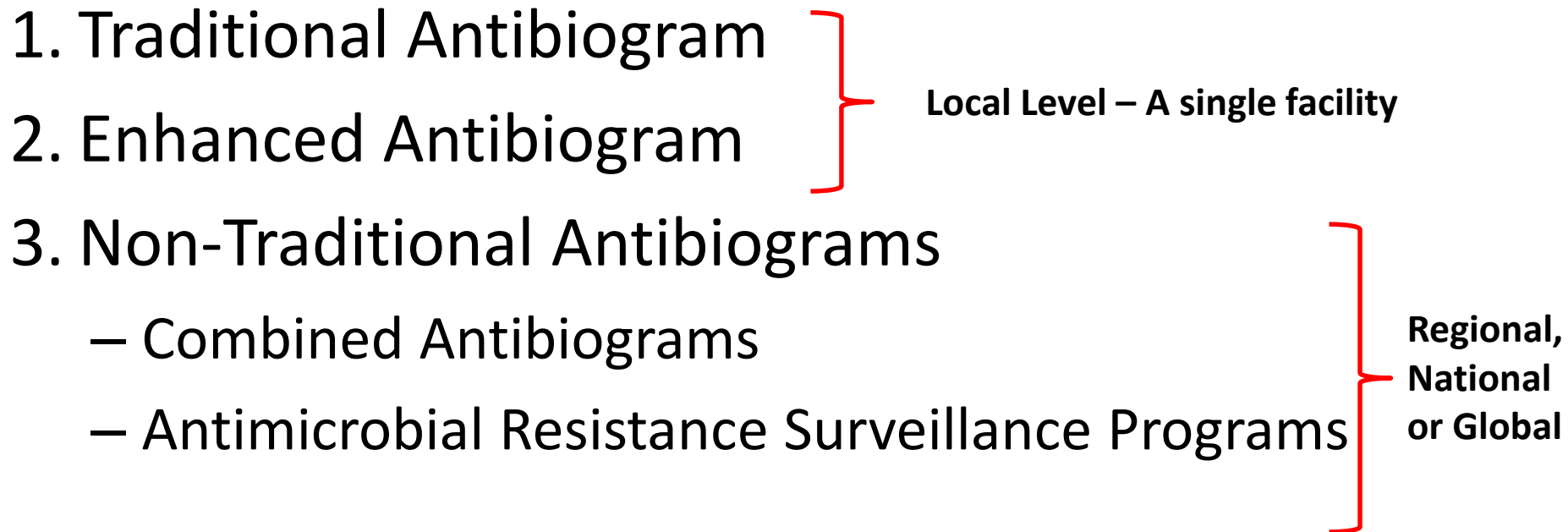
Terminology

- Sensitive
 - Based on the pharmaco-dynamics of an antimicrobial agent administered according to the normally recommended dosage and the organism causing an infection, the agent *will most likely inhibit the organism in vivo*.
- Intermediate (indeterminate)/Susceptible Dose Dependent
 -*might inhibit the organism in vivo*. /Use higher dose
 -
- Resistant
 -*will most likely not inhibit the organism in vivo*.
- Non-Susceptible
 -*Not enough data to know if it is likely to inhibit the organism in vivo*.
- Epidemiological Cutoff Values
 - Determines the MIC of Wildtype and non-WT.

Where does the data come from?

- Microbiology AST instruments
- Microbiology LIS
- Electronic Health Records (EHR)
- Clinical decision support system (CDSS)

3 Types of Cumulative AST Data Reports

1. Traditional Antibigram
 2. Enhanced Antibigram
 3. Non-Traditional Antibigrams
 - Combined Antibigrams
 - Antimicrobial Resistance Surveillance Programs
- Local Level – A single facility
- Regional, National or Global
- 

What Is An Antibigram?

- Presentation of cumulative antimicrobial susceptibility testing (AST) data from a single institution on an annual basis

Appendix E1. Cumulative Antimicrobial Susceptibility Report Example – Antimicrobial Agents Listed Alphabetically (Hypothetical Data)

Memorial Medical Center
1 January – 31 December 2012 Cumulative Antimicrobial Susceptibility Report*
Percent Susceptible

Gram-Negative Organisms	No. Strains	Aminiacin	Ampicillin	Cefazolin	Cefotaxime	Ceftazidime	Ciprofloxacin	Nitrofurantoin†	Gentamicin	Meropenem	Piperacillin-tazobactam	Trimethoprim-sulfamethoxazole	Tobramycin
<i>Acinetobacter baumannii</i>	32	80	R	R	34	52	51	–‡	60	80	46	58	59
<i>Citrobacter freundii</i>	49	100	R	R	72	67	90	78	100	99	67	67	100
<i>Enterobacter aerogenes</i>	31	100	R	R	68	69	92	85	91	99	74	95	91
<i>Enterobacter cloacae</i>	76	99	R	R	61	62	92	81	90	99	77	84	90
<i>Escherichia coli</i>	1433	99	36	68	96	94	72	98	91	99	51	65	92
<i>Klebsiella pneumoniae</i>	543	99	R	72	91	92	84	74	94	95	86	81	94
<i>Morganella morganii</i>	44	100	R	R	85	81	99	R	100	99	64	75	100
<i>Proteus mirabilis</i>	88	100	87	80									
<i>Pseudomonas aeruginosa</i>	397	97	R	R									
<i>Salmonella</i> spp.	32	–	88	–									
<i>Serratia marcescens</i>	50	100	R	R									
<i>Shigella</i> spp.	33	–	64	–									
<i>Stenotrophomonas maltophilia</i>	72	R	R	R	R	63	6	R	R	R	–	98	R

“Routine” Cumulative antibiogram
Generally...all isolates from a facility

* The percent susceptible for each organism/antimicrobial combination was generated by including the first isolate of that organism encountered on a given patient.

† Nitrofurantoin data from testing urine isolates only.

‡ (–) drug not tested or drug not indicated.

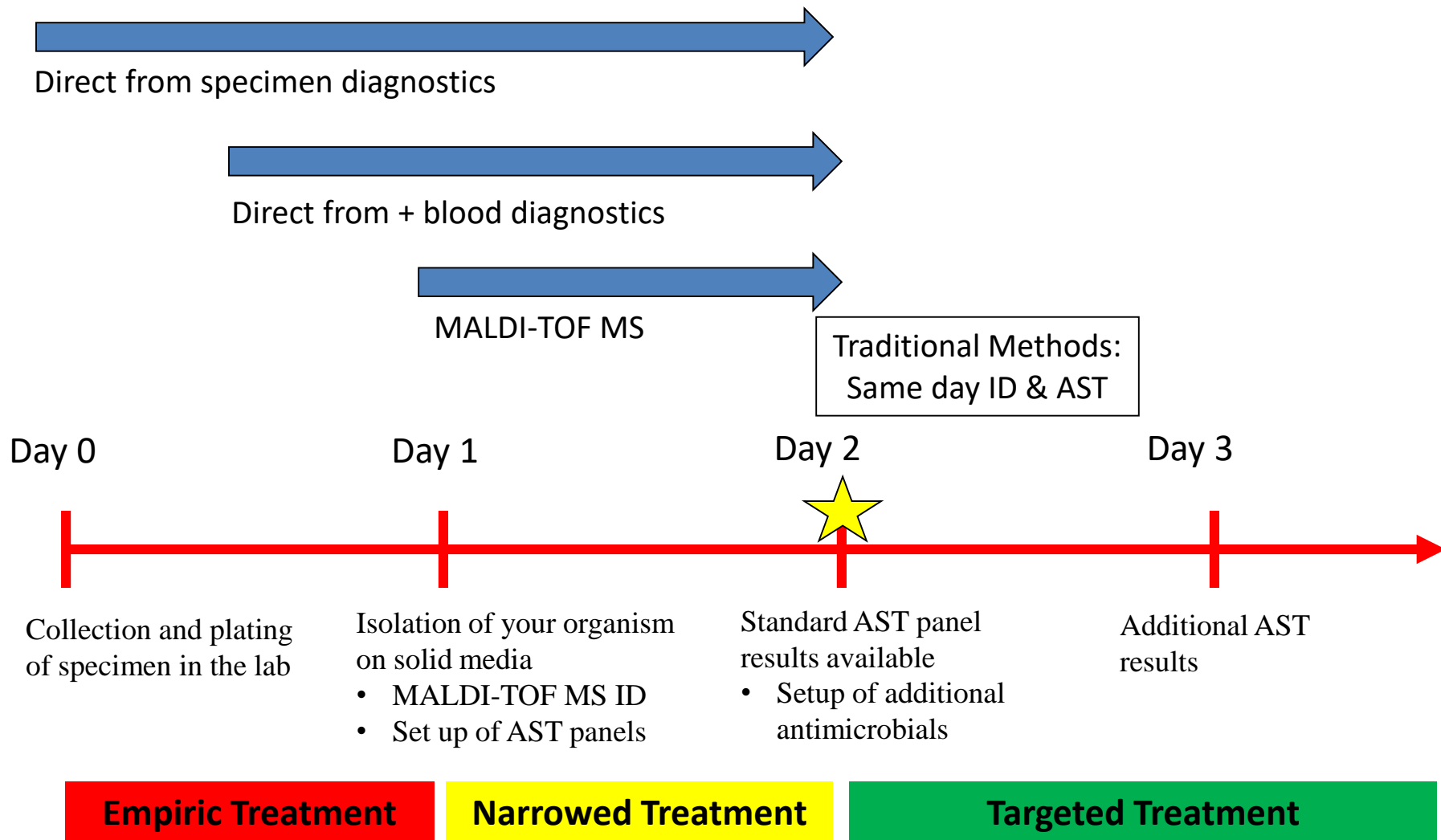
Abbreviations: No., number; R, intrinsic resistance.

The **Why**? - Purpose of the Antibigram

- To help clinicians choose initial empiric therapy
- Many more applications
 - Dr. Kim Claeys presentation on February 6th discussed using the antibiogram for Antimicrobial Stewardship applications

Importance & Reliance on Antibigrams Grow!

Courtesy of Trish Simner

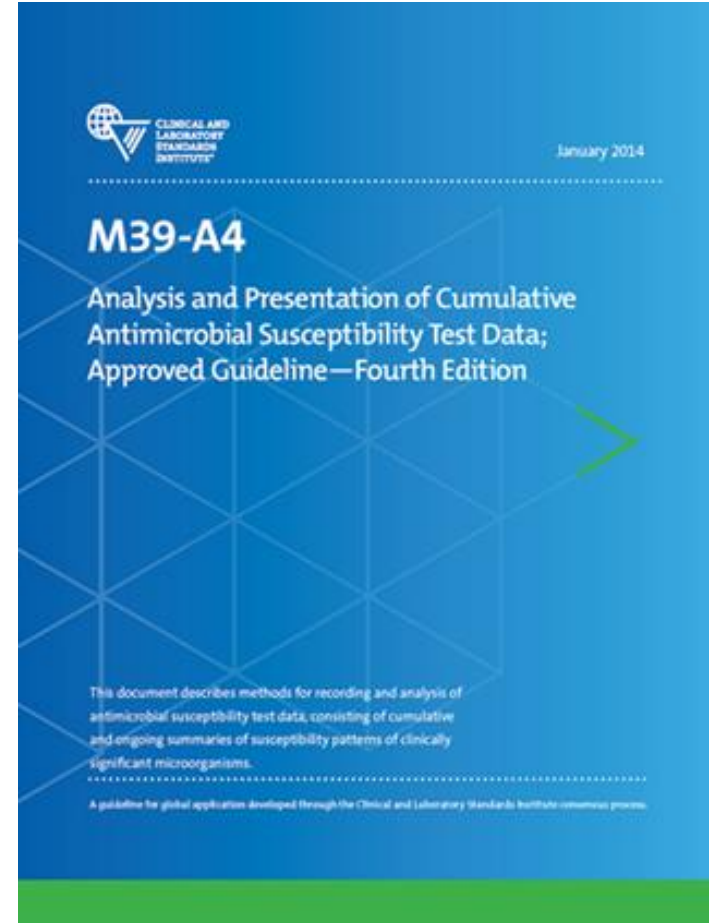


Who is Responsible for Creating the Antibigram?

- Traditionally the microbiology laboratory
 - Driven by access to the data from AST instruments or the LIS
- Shifting towards stronger collaborations with Antimicrobial Stewardship Programs
 - Automated EHR based antibiograms
- Should be a collaborative effort
 - Clinical microbiologists, pharmacists, physicians, IT specialists

Where Do You Start?

- M39-A4: Analysis and Presentation of Cumulative Antimicrobial Susceptibility Test Data; Approved Guideline-Fourth Edition
- M39-A5 currently being worked on. 2019-2020
 - A newly created section on LTC



Preparation of Cumulative Antibigram Recommendations

- **WHEN?**-Analyze/present data at least **annually**
- Include only verified **final results**
- Include only species with **≥ 30** isolates
- Include **diagnostic** (not surveillance) isolates
- Include the **1st isolate/patient**; no duplicate isolates
- Only include **routinely tested** antimicrobial agents
- Report only **%S** and do not include I%

The Cumulative Antibigram Report

- Analyzes data from **routine antimicrobial susceptibility tests** performed in the clinical laboratory
- Separate report prepared for each **healthcare facility**
- Primarily used to guide **empiric therapy**
- Sometimes used to **monitor resistance**
 - Changes in %S from year to year
- **Highly impacted** by
 - patient population served
 - culturing practices
 - If cultures only performed when patients fail therapy
 - Laboratory antimicrobial susceptibility testing and reporting policies
 - Temporal outbreaks

Organism Specific Recommendations

Bug/Drug	Presentation of Data
<i>Streptococcus pneumoniae</i> and penicillin	List the %S using oral, meningitis and non-meningitis breakpoints
<i>Streptococcus pneumoniae</i> and cefotaxime, ceftriaxone, cefepime	List the %S using meningitis and nonmeningitis breakpoints
Viridans group streptococci and penicillin	List both the %S and %I
<i>Staphylococcus aureus</i>	List %S for all isolates and the methicillin-resistant <i>S. aureus</i> (MRSA) subset
<i>E. coli</i> , <i>K. pneumoniae</i> and <i>P. mirabilis</i> and cefazolin	List % S using urine and non-urine breakpoints

Stratification of AntibioGrams

- Nursing site or site of care
 - ICU, burn unit, ED, outpatient clinic
- Specimen type of infection site
 - Urine, blood
- Clinical service or patient population
 - Surgical, pediatric, transplant, cancer



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Answers to Commonly Asked Questions

How do I Apply Intrinsic Resistance?

B1. Enterobacteriaceae

Antimicrobial Agent Organism	Ampicillin	Amoxicillin-clavulanate	Ampicillin-sulbactam	Piperacillin	Ticarcillin	Cephalosporins I: Cefazolin, Cephalothin	Cephameycins: Cefoxitin, Cefotetan	Cephalosporin II: Cefuroxime	Imipenem	Tetracyclines	Tigecycline	Nitrofurantoin	Polymyxin B Colistin	Aminoglycosides
<i>Citrobacter freundii</i>	R	R	R			R	R	R						
<i>Citrobacter koseri</i>	R			R	R									
<i>Enterobacter cloacae</i> complex ^a	R	R	R			R	R	R						
<i>Escherichia coli</i>	There is no intrinsic resistance to β -lactams in this organism.													
<i>Escherichia hermannii</i>	R				R									
<i>Hafnia alvei</i>	R	R	R			R	R							
<i>Klebsiella</i> (formerly <i>Enterobacter</i>) <i>aerogenes</i>	R	R	R			R	R	R						
<i>Klebsiella pneumoniae</i>	R				R									
<i>Morganella morganii</i>	R	R				R		R	^b		R	R	R	
<i>Proteus mirabilis</i>	There is no intrinsic resistance to penicillins and cephalosporins in this organism.								^b	R	R	R	R	
<i>Proteus penneri</i>	R					R		R	^b	R	R	R	R	
<i>Proteus vulgaris</i>	R					R		R	^b	R	R	R	R	
<i>Providencia rettgeri</i>	R	R				R			^b	R	R	R	R	
<i>Providencia stuartii</i>	R	R				R			^b	R	R	R	R	^c

The most up-to-date Intrinsic Resistance tables are located in the current M100 document.

What Do You Do With Susceptible Dose Dependent (SDD) Results?

- SDD: an interpretive category defined by a breakpoint that susceptibility of an isolate is dependent on the dosing regimen that is used in the patient
 - Cefepime and *Enterobacteriaceae*
 - Fluconazole and *C. albicans*, *C. glabrata*, *C. parapsilosis*, & *C. tropicalis*
 - New in 2019:
 - Daptomycin and *Enterococcus* spp
 - Ceftaroline and *Staphylococcus aureus*
 - Report both % S & %SDD either in the Table or as a footnote

	N	% S Cefepime	%SDD Cefepime
<i>Escherichia coli</i>	574	92 ^a	3
<i>Klebsiella pneumoniae</i>	132	84 ^b	2

^a: “In addition, to the 92% S results, 3% were SDD (MIC 4 to 8 µg/mL) and 5% were R (MIC >16 µg/mL) to cefepime”

Why Do We Need a Minimum of N=30?

- Less statistical validity of data
 - Small numbers can skew the data

How Reliable is a Report of 80% Susceptible for *E. coli* and Ciprofloxacin?

Sample size	% S (95% CI)
10	44 to 97
100	71 to 87
1000	77 to 82

What Do I Do If We Don't Reach $N \geq 30$?

- So what can you do?
 - Analyze multiple years – add footnote
 - Report the results from $N < 30$ with a footnote
 - “Calculated from fewer than the standard recommendation of 30 isolates”
 - Group several species within a genus together
 - Aggregate data from multiple smaller facilities with a similar patient population in the same geographic area

Discuss more LTCF specific later



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Enhanced Antibigram

What Are Enhanced Antibigrams?

- Segregating cumulative antibiogram data by one or more of the following:
 - **Location** – e.g., Inpatient vs Outpatient or ICU vs Oncology vs Non-ICU/Non-Oncology Wards
 - **Specimen type** – e.g. urine or blood specific
 - **Clinical condition** – e.g. cystic fibrosis, burn patients
 - **Patient Age** – e.g., pediatrics vs adults
 - **Resistance Phenotype** – e.g., MRSA, MSSA, carbapenem-resistant *Enterobacteriaceae*
 - **Organism** – e.g. anaerobe antibiogram
 - **ASP Antibigrams** – e.g. novel agents or last resort agents (colistin)
- Resistance Profiles
 - % Susceptible for combinations of drugs
 - % Susceptible for groups of organisms (e.g., all GNR from blood)

Combination of Antimicrobial Agents

Organism	No. Strains	% Susceptible							
		CIP	CTZ	IMP	TOB	CTZ and/or CIP	IMP and/or CIP	CTZ and/or TOB	IMP and/or TOB
<i>Pseudomonas aeruginosa</i>	814	69	80	79	86	86	84	91	91

Organisms resistance characteristics

Organism	No. Strains	% Susceptible								
		CLI	DOX	ERY	GEN	OXA	PEN	RIF	SXT	VAN
All <i>S. aureus</i>	1317	80	98	50	93	68	13	98	96	100
Oxacillin-resistant <i>S. aureus</i> (MRSA)	449	44	96	4	79	0	0	95	94	100
Oxacillin-susceptible <i>S. aureus</i> (MSSA)	904	97	99	72	99	100	18	99	97	100

Organism	No. Strains	% Susceptible									
		AMK	AMP	CFZ	CRO	CIP	GEN	IMP	PTZ	TET	SXT
<i>K. pneumoniae</i> (all)	1163	63	-	44	48	46	74	64	53	84	46
<i>K. pneumoniae</i> (ESBL-producing)	233	30	-	0	0	6	48	100	0	84	3
<i>K. pneumoniae</i> (KPC-producing)	361	5	-	0	0	0	28	0	0	82	0
<i>K. pneumoniae</i> (non-ESBL or KPC-producing)	569	100	-	84	99	94	96	100	88	87	95



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Non-Traditional Antibiofilms

What About Non-Traditional Antibigrams?

- Accumulate AST data outside of a single institution
 - Combined Regional Antibigrams
 - Antimicrobial Resistance Surveillance Programs (ARSP)
 - Creating an ARSP report – New in M39-A5

Characteristic	Routine Antibigram	Non-Traditional Antibigram
Study Period	Annually	Defined by study
# of Institutions	One	Multiple
Presentation	Table	Report with I, M&M, R and D

When To Consider Utilizing Non-Traditional Antibigram Data?

- The use of a local cumulative antibiogram **is preferred to guide initial empiric therapy**
- Non-Traditional antibiogram data:
 - Used when local AST data are not available, are limited in size or scope
 - Used as a benchmark to compare local data to regional and national findings

Combined REGIONAL Antibigram

- Compilation of data from facility-level antibiograms
- Susceptibility was defined by local labs in all circumstances
- Created a report with an Introduction, Methodology Notes, Antibigram Table & Breakdown by Individual Organisms
- Methodology Notes Included:
 - Differences in breakpoints (eg cephalosporin & carbapenem breakpoints)
- Differences in agents within a class (eg ciprofloxacin vs levofloxacin)



2015 LOS ANGELES COUNTY ACUTE CARE HOSPITAL ANTI-BIOTIC RESISTANCE REPORT
Gram-Negative Organisms



		Penicillins		Cephalosporins			Carbapenems		Aminoglycosides			Quinolone	Other
Percent Susceptible (Number of isolates tested)	# of all isolates tested (n = all isolates reported)	Ampicillin/ Sulbactam	Piperacillin/ Tazobactam	Ceftriaxone	Cefazidime	Cefepime	Ertapenem	Meropenem	Amikacin	Gentamicin	Tobramycin	Ciprofloxacin/ Levofloxacin	Trimethoprim/ Sulfamethoxazole
<i>Acinetobacter</i> sp.	3189 (66)	-	33 (1,873)	11 (1,475)	30 (2,184)	34 (1,864)	R	53 (1,561)	43 (2,004)	41 (2,970)	46 (2,126)	33 (3,024)	49 (2,859)
<i>Citrobacter freundii</i>	1975 (43)	R	97 (1,823)	82 (1,889)	83 (1,503)	98 (1,713)	99 (1,156)	99 (1,142)	100 (1,536)	92 (1,924)	93 (1,138)	91 (1,975)	81 (1,939)
<i>Citrobacter koseri</i>	631 (23)	-	99 (631)	96 (631)	97 (427)	100 (456)	100 (223)	100 (184)	99 (389)	99 (631)	99 (428)	99 (631)	96 (601)
<i>Enterobacter</i> sp.	8122 (66)	R	82 (7,507)	80 (7,307)	82 (6,204)	96 (7,040)	96 (4,417)	99 (4,636)	100 (6,235)	97 (7,972)	96 (4,630)	96 (8,120)	92 (8,018)
<i>Escherichia coli</i>	139212 (73)	55 (25,534)	93 (115,257)	86 (105,020)	86 (95,157)	86 (90,175)	100 (78,427)	100 (84,318)	99 (104,151)	86 (129,487)	81 (67,956)	70 (129,130)	66 (123,819)
<i>Klebsiella</i> sp.	30655 (72)	-	84 (25,586)	86 (23,006)	86 (19,120)	85 (19,895)	98 (15,578)	97 (17,025)	94 (22,223)	91 (27,834)	82 (16,128)	86 (28,047)	82 (26,934)
<i>Morganella</i> sp.	2235 (52)	-	96 (2,233)	88 (2,055)	81 (1,811)	98 (1,921)	100 (1,148)	100 (1,127)	99 (1,913)	71 (2,234)	66 (1,358)	60 (2,231)	55 (2,154)
<i>Proteus</i> sp.	16908 (68)	-	98 (15,836)	90 (15,682)	92 (13,067)	92 (13,832)	99 (9,018)	99 (9,903)	99 (13,470)	83 (16,554)	84 (10,176)	68 (16,738)	68 (16,491)
<i>Providencia</i> sp.	1618 (16)	-	73 (1,542)	66 (1,404)	55 (1,315)	77 (1,285)	88 (228)	90 (553)	91 (1,442)	11 (1,299)	14 (960)	11 (1,512)	46 (1,513)
<i>Pseudomonas aeruginosa</i>	22804 (73)	R	83 (10,040)	R	82 (18,315)	84 (19,015)	R	82 (14,261)	95 (19,491)	83 (22,271)	91 (19,850)	69 (22,132)	R
<i>Serratia</i> sp.	2676 (58)	R	91 (2,098)	90 (2,403)	91 (2,188)	97 (2,203)	97 (1,414)	98 (1,579)	97 (2,188)	97 (2,757)	85 (1,677)	88 (2,646)	97 (2,544)
<i>Stenotrophomonas maltophilia</i>	1719 (50)	R	R	R	37 (848)	R	R	R	R	R	R	79 (1,052)	90 (1,548)

Data not collected denoted by "-".
R, intrinsic resistance

<http://publichealth.lacounty.gov/acd/AntibiogramData.htm>



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Applications of Cumulative AST Data

Many Applications of Cumulative AST data

Stake Holder	Application
Physicians	Empiric therapy decisions
Clinical Microbiology Laboratories	Benchmarking, quality control, role of rapid diagnostics
Antimicrobial Stewardship Programs	Antimicrobial therapy recommendations and formulary decisions
Infection Prevention and Control	Benchmarking to evaluate infection control practices
Pharmaceutical Industry	Informs new drug development
Regulatory	Informs regulatory practices
Public Health	Monitoring changes in resistance levels and public health interventions



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Increasing Awareness of Antibiograms

Increasing Awareness of Antibigram Data

Methods for Antibigram Data Dissemination

Pocket guides/booklets

Laminated posters

Hospital newsletter article

Posting within hospital intranet/EMR

Email to all prescribers

Smartphone or tablet applications

Presentations

Increasing Awareness of Antibiogram Data

Appendix G – provides stepwise instructions on presenting the local cumulative antibiogram data to healthcare professionals

- Explain purpose of the local cumulative antibiogram with a brief description of how the report is prepared
- Describe any software limitations
- Describe the rationale used for separating data into subgroups for the report
- Present graphs and charts for trends that are monitored each year



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Recommendations in the New M39-A5 for LTC

Optimizing Culturing Practices in LTCF

- *Suspected Urinary Tract Infection* – To avoid over-culturing, consider developing a policy with the LTCF reference lab to determine if culture can be performed ONLY on urine specimens with significant pyuria (auto-reflex to culture).
- *Suspected Pneumonia* – Obtain an expectorated sputum sample, if possible, for Gram stain and culture.
- *Suspected Skin and Soft Tissue Infection* – If the skin infection is associated with an abscess or area of purulence, send a sample of the pus to the lab for culture.

Responsibility for Cumulative Antibigram Development

- The willingness of the referral lab to either develop the antibiogram or provide susceptibility reports for antibiogram development should be determined.
 - Guidelines that will be followed for antibiogram development (e.g., CLSI M39)
 - Information (e.g., bacteria, antibiotics, etc.) that should be included in the antibiogram
 - Method for collection of cumulative susceptibility data
 - Method for data analysis, presentation and formatting (e.g., time period of antibiogram, data segregation techniques, the utility of infection-specific reports, etc)
- Multiple Referral Labs
 - Variations in laboratory practice must be considered (breakpoints)
 - Data formatted the same way
 - Appropriate and correct data from each laboratory

Data Analysis Techniques

- First Isolate per Patient
 - First isolate per reporting period
- Handling of small numbers (≤ 30)
 - Consider combining data from multiple years
 - Consider combining species, if applicable
 - Consider using data from other sources
 - Evaluate current culturing practices to assure that all patients with suspected infection are being cultured appropriately.
 - Consider constructing a cumulative antibiogram from patients in the general community in age category 65 and older.

More information and resources on the development of a LTCF Antibigram

- <https://www.ahrq.gov/nhguide/toolkits/help-clinicians-choose-the-right-antibiotic/index.html>

Summary

- M39-A4 provides guidelines for creating a cumulative antibiogram
- There are 3 types of cumulative antibiograms
- Cumulative antibiograms can be stratified by different patient, hospital, or organism characteristics
- New Guidelines specific for LTCF will be in the new edition of M39-A5 coming out in 2020